

RESEARCH ARTICLE

Open Access



Serum tissue inhibitor of matrix metalloproteinase-1 levels are associated with mortality in patients with malignant middle cerebral artery infarction

Leonardo Lorente^{1*}, María M. Martín², Luis Ramos³, Juan J. Cáceres⁴, Jordi Solé-Violán⁵, Mónica Argueso⁶, Alejandro Jiménez⁷, Juan M. Borreguero-León⁸, Josune Orbe⁹, José A. Rodríguez⁹ and José A. Páramo⁹

Abstract

Background: In the last years, circulating matrix metalloproteinases (MMP)-9 levels have been associated with functional outcome in ischemic stroke patients. However the prognostic value of circulating levels of tissue inhibitor of matrix metalloproteinases (TIMP)-1 and MMP-10 in functional outcome of ischemic stroke patients has been scarcely studied. In addition, to our knowledge, serum MMP-9, MMP-10 and TIMP-1 levels in patients with malignant middle cerebral artery infarction (MMCAI) for mortality prediction have not been studied, and these were the objectives of this study.

Methods: This was a multicenter, observational and prospective study carried out in six Spanish Intensive Care Units. We included patients with severe MMCAI defined as Glasgow Coma Scale (GCS) lower than 9. We measured circulating levels of MMP-9, MMP-10, TIMP-1, in 50 patients with severe MMCAI at diagnosis and in 50 healthy subjects. Endpoint was 30-day mortality.

Results: Patients with severe MMCAI showed higher serum levels of MMP-9 ($p = 0.001$), MMP-10 ($p < 0.001$), and TIMP-1 ($p = 0.02$) than healthy subjects. Non-surviving MMCAI patients ($n = 26$) compared to survivor ones ($n = 24$) showed higher circulating levels of TIMP-1 ($p < 0.001$), MMP-10 ($p = 0.02$) and PAI-1 ($p = 0.02$), and lower MMP-9 levels ($p = 0.04$). Multiple binomial logistic regression analysis showed that serum TIMP-1 levels > 239 ng/mL are associated with 30-day mortality (OR = 5.82; 95 % CI = 1.37-24.73; $P = 0.02$) controlling for GCS and age. The area under the curve for TIMP-1 as predictor of 30-day mortality was 0.81 (95 % CI = 0.67-0.91; $P < 0.001$). We found an association between circulating levels of TIMP-1 and MMP-10 ($\rho = 0.45$; $P = 0.001$), plasminogen activator inhibitor (PAI)-1 ($\rho = 0.53$; $P < 0.001$), and tumor necrosis factor (TNF)-alpha ($\rho = 0.70$; $P < 0.001$).

Conclusions: The most relevant and new findings of our study, were that serum TIMP-1 levels in MMCAI patients were associated with mortality, and could be used as a prognostic biomarker of mortality in MMCAI patients.

Keywords: TIMP-1, Ischemic stroke, Patients, Mortality, Injury

* Correspondence: lorentemartin@msn.com

¹Intensive Care Unit, Hospital Universitario de Canarias, Ofra, s/n. La Laguna, 38320 Santa Cruz de Tenerife, Spain

Full list of author information is available at the end of the article

Background

Ischemic stroke is an important cause of disability, mortality and resources consume [1]. Matrix metalloproteinases (MMPs) are implicated in degradation and remodelling of the extracellular matrix (ECM). That family of zinc-containing endoproteinases can be classified according to the substrate specificity as collagenases (MMP-1, -8 and -13), gelatinases (MMP-2 and -9), stromelysins (MMP-3, -10, -11), matrilysins (MMP-7), elastases (MMP-12) and membrane-type (MT-MMPs, MMP-14, -15, -16 and -17). MMP activity is regulated by specific tissue inhibitors of matrix metalloproteinases (TIMPs). MMPs are involved in physiological functions such as morphogenesis, menstrual cycle, tissue remodeling and angiogenesis; and also in some diseases with abnormal ECM turnover, such as arthritis, sepsis, tumour invasion and atherosclerosis [2–7].

In the last years, MMPs have been found to play a role in cerebral ischemia [8–10]. In some studies higher circulating MMP-9 levels were found in ischemic stroke patients than in controls [11–15], and in ischemic stroke patients with worse functional outcome [11–19]. However the prognostic value of circulating levels of TIMP-1 [20] and MMP-10 [21] in functional outcome of ischemic stroke patients has been scarcely studied. Circulating TIMP-1 levels have been associated with poor prognosis in a community-based cohort of elderly men risk [22], patients with coronary artery disease [23], and in different cancer types, such as lung [24] breast [25] colorectal [26] and gastric cancer [27]. There have been found higher TIMP-1 concentrations in infarcted brain tissue compared to healthy cerebral areas [28], higher expression of TIMP-1 in monocytes of ischemic stroke patients than in healthy controls [29], and higher circulating TIMP-1 levels in ischemic stroke patients than in healthy controls [30–33]. In addition, there has been found an association between serum TIMP-1 levels and mortality in patients with severe trauma brain injury [34].

To our knowledge, serum MMP-9, MMP-10 and TIMP-1 levels in patients with malignant middle cerebral artery infarction (MMCAI) for mortality prediction have not been studied, and these were the objectives of this study.

Methods

Design and subjects

This is a multicenter, observational, prospective study carried out in 6 Intensive Care Units of Spain. The study was approved by the Institutional Review Board of the 6 participant hospitals: Hospital Universitario de Canarias (La Laguna, Santa Cruz de Tenerife, Spain), Hospital Universitario Nuestra Señora de Candelaria (Santa Cruz de Tenerife, Spain), Hospital General de La Palma (La Palma, Spain), Hospital Clínico Universitario de Valencia (Valencia, Spain), Hospital Insular (Las Palmas de Gran Canaria,

Spain), Hospital Universitario Dr. Negrín (Las Palmas de Gran Canaria, Spain). The written informed consent from the patients or from their legal guardians was obtained.

We included 50 patients with severe MMCAI and 50 healthy volunteer control subjects. Severity of MMCAI was classified according to Glasgow Coma Scale (GCS) [35], and severe was defined as $GCS \leq 8$. Exclusion criteria were: age less than 18 years, inflammatory or malignant disease.

Variables recorded

The following variables were recorded for each patient: sex, fibrinolytic therapy, decompressive craniectomy, age, temperature, sodium, glycemia, leukocytes, pressure of arterial oxygen (PaO₂), PaO₂/ pressure of arterial oxygen/ fraction inspired oxygen (FI_{O₂}) ratio, bilirubin, creatinine, hemoglobin, GCS, lactic acid, platelets, international normalized ratio (INR), activated partial thromboplastin time (aPTT), fibrinogen, Acute Physiology and Chronic Health Evaluation II (APACHE II) score [36]. The end-point of the study was 30-days mortality.

Blood sample collection

Blood samples of 50 patients with severe MMCAI were collected at the moment of the diagnosis and of 50 controls to measure the concentrations of MMP-9, MMP-10, TIMP-1, tumor necrosis factor (TNF)-alpha, and plasminogen activator inhibitor (PAI)-1. To avoid the possible dispersion of serum level results, all the samples were processed at same time and in the same laboratory, at the end of the recruitment process.

Determination of serum MMP-9, MMP-10, TIMP-1 and TNF-alpha levels

Serum separator tubes were used to determine serum MMP-9, MMP-10, TIMP-1 and TNF-alpha levels. Venous blood samples were taken and centrifuged within 30 min at 1000 g for 15 min, and the serum was removed and frozen at -80 °C until measurement.

MMP-9, MMP-10 and TIMP-1 assays were performed at the Atherosclerosis Research Laboratory of CIMA-University of Navarra (Pamplona, Spain) and were assayed by specific ELISAs (Quantikine®, R&D Systems, Abingdon, United Kingdom) according to the manufacturer's instructions with a serum dilution of 1:80, 1:2 and 1:100 respectively. The interassay coefficients of variation (CV) were <8 % (n = 20) and detection limit for the assays were 0.31 ng/ml, 78.1 pg/ml and 0.15 ng/ml respectively.

TNF-alpha serum levels were measured in the Laboratory Department of the Hospital Universitario de Canarias (La Laguna, Santa Cruz de Tenerife, Spain) by a solid-phase, chemiluminiscent immunometrics assays kit (Immulite®, Siemens Healthcare Diagnostics Products, Llanberis, United Kingdom); and the interassays CV was <6.5 % (n = 20) and detection limit for the assay was 1.7 pg/mL.

Determination of plasma PAI-1 levels

Venous blood samples were collected in citrate collected plasma tubes and centrifuged within 30 min at 1000*g for 15 min. The plasma was removed and frozen at -80 °C until measurement. PAI-1 assay was performed at the Laboratory Department of the Hospital Universitario de Canarias (La Laguna, Santa Cruz de Tenerife, Spain). PAI-1 antigen levels were assayed by specific ELISA (Imubind Plasma PAI-1 American Diagnostica, Inc, Stanford, CT, USA). The interassay CV of PAI-1 assay was <5 % (n = 20) and detection limits was 1 ng/mL.

Statistical methods

Continuous variables are reported as medians and interquartile ranges. Categorical variables are reported as frequencies and percentages. Comparisons of continuous variables between groups were carried out using Wilcoxon-Mann-Whitney test. Comparisons between groups on categorical variables were carried out with chi-square test.

Multiple binomial logistic regression analysis was applied to determine the independent contribution of TIMP-1 on 30-day mortality, controlling for GCS and age. Odds Ratio and 95 % confidence intervals were calculated as measurement of the clinical impact of the predictor variables.

Receiver operating characteristic (ROC) analysis was carried out to determine the goodness-of-fit of the of serum TIMP-1 levels to predict 30-day mortality. Kaplan-Meier analysis of survival at 30 days and comparisons by log-rank test were carried out using serum TIMP-1 levels lower/higher than 239 ng/mL as the independent variable and survival at 30 days as the dependent variable. The association between continuous variables was carried out using Spearman's rank correlation coefficient. A *P* value of less than 0.05 was considered statistically significant. Statistical analyses were performed with SPSS 17.0 (SPSS Inc., Chicago, IL, USA) and NCSS 2000 (Kaysville, Utah) and LogXact 4.1, (Cytel Co., Cambridge, MA).

Table 1 Characteristics of healthy controls and patients with severe MMCAI

	Healthy controls (n = 50)	Patients (n = 50)	p-value
Gender female - n (%)	13 (26.0 %)	17 (34 %)	0.51
Age (years) - median (p 25-75)	57 (50-63)	60 (51-69)	0.11
TIMP-1 (ng/mL) - median (p 25-75)	226 (213-241)	261 (199-387)	0.02
MMP-9 (ng/mL) - median (p 25-75)	498 (350-735)	749 (488-1200)	0.001
MMP-10 (pg/mL) - median (p 25-75)	466 (288-614)	1027 (556-1409)	<0.001

MMP = matrix metalloproteinase; TIMP = tissue inhibitor of matrix metalloproteinases

Table 2 Clinical and biochemical characteristics of survivor and non-survivor MMCAI patients

	Survivors (n = 24)	Non-survivors (n = 26)	P value
Gender female - n (%)	8 (33.3)	9 (34.6)	0.99
Decompressive craniectomy - n (%)	7 (29.2)	5 (19.2)	0.51
Age (years) - median (p 25-75)	47 (32-67)	66 (45-76)	0.14
Temperature (°C) - median (p 25-75)	36.5 (35.7-37.0)	37.0 (35.7-37.8)	0.26
Sodium (mEq/L)- median (p 25-75)	140 (138-145)	140 (137-146)	0.91
Glycemia (g/dL) - median (p 25-75)	133 (105-170)	135 (110-154)	0.92
Leukocytes-median*10 ³ / mm ³ (p 25-75)	12.8 (9.8-16.9)	14.4 (11.9-21.9)	0.49
PaO ₂ (mmHg) - median (p 25-75)	110 (101-194)	104 (85-139)	0.10
PaO ₂ /FIO ₂ ratio - median (p 25-75)	246 (192-327)	248 (175-320)	0.41
Bilirubin (mg/dl) - median (p 25-75)	0.50 (0.38-0.90)	0.53 (0.30-1.20)	0.76
Creatinine (mg/dl) - median (p 25-75)	0.80 (0.60-1.10)	1.01 (0.85-1.45)	0.052
Hemoglobin (g/dL) - median (p 25-75)	12.0 (11.3-13.8)	12.0 (11.0-15.1)	0.92
GCS score - median (p 25-75)	7 (6-8)	6 (4-8)	0.10
Lactic acid (mmol/L)-median (p 25-75)	1.25 (0.93-1.68)	1.50 (1.01-3.15)	0.08
Platelets - median*10 ³ /mm ³ (p 25-75)	227(183-308)	152 (123-190)	0.003
INR - median (p 25-75)	1.07 (1.01-1.20)	1.20 (1.07-1.48)	0.16
aPTT (seconds) - median (p 25-75)	28 (25-29)	26 (25-33)	0.96
Fibrinogen (mg/dl) - median (p 25-75)	440 (335-494)	409 (322-598)	0.71
APACHE-II score - median (p 25-75)	20 (16-25)	22 (19-29)	0.14
MMP-9 (ng/mL) - median (p 25-75)	963 (731-1218)	672 (384-1088)	0.04
MMP-10 (pg/mL) - median (p 25-75)	785 (550-1114)	1264 (608-1759)	0.02
TIMP-1 (ng/mL) - median (p 25-75)	204 (172-264)	343 (240-493)	<0.001
PAI-1 (ng/mL) - median (p 25-75)	24.0 (19.3-40.8)	51.5 (28.3-95.3)	0.02
TNF-alpha (pg/mL) - median (p 25-75)	9.25 (9.02- 10.63)	12.95 (10.03- 15.08)	0.01

P 25-75 = percentile 25th-75th; PaO₂ = pressure of arterial oxygen/fraction inspired oxygen; FIO₂ = pressure of arterial oxygen/fraction inspired oxygen; GCS = Glasgow Coma Scale; ISS = Injury Severity Score; INR = international normalized ratio; aPTT = activated partial thromboplastin time; APACHE II = Acute Physiology and Chronic Health Evaluation; MMP = matrix metalloproteinase; TIMP = tissue inhibitor of matrix metalloproteinases; PAI = plasminogen activator inhibitor; TNF = tumor necrosis factor

Results

Patients with severe MMCAI showed higher serum levels of MMP-9, MMP-10 and TIMP-1 than healthy subjects (Table 1).

We found that non-surviving MMCAI patients (n = 26) compared to survivors ones (n = 24) showed higher circulating levels of MMP-10, TIMP-1, PAI-1 and TNF-alpha, and lower MMP-9 levels (Table 2).

Multiple binomial logistic regression analysis showed that serum TIMP-1 levels > 239 ng/mL are associated with 30-day mortality (OR = 5.82; 95 % CI = 1.37-24.73; P = 0.02) controlling for GCS and age (Table 3).

The area under the curve (AUC) for TIMP-1 as predictor of 30-day mortality was 0.81 (95 % CI = 0.67-0.91; P < 0.001) (Fig. 1).

Survival analysis showed that patients with serum TIMP-1 higher than 239 ng/mL presented higher 30-day mortality than patients with lower levels (Hazard ratio = 3.6; 95 % CI = 1.67-7.82; P = 0.004) (Fig. 2).

We found an association between circulating levels of TIMP-1 and MMP-10 (rho = 0.45; P = 0.001), PAI-1 (rho = 0.53; P < 0.001), and TNF-alpha (rho = 0.75; P < 0.001).

Discussion

The novel findings of our study were the following: a) non-surviving severe MMCAI patients had higher serum TIMP-1 and MMP-10 levels than surviving patients; b) there is an association between circulating levels of TIMP-1, PAI-1, and TNF-alpha in patients with severe MMCAI; c) serum TIMP-1 levels could be used as prognostic biomarker in patients with severe MMCAI.

We found that patients with severe MMCAI showed higher serum levels of MMP-9, MMP-10 and TIMP-1 than healthy subjects. Previously there were found higher circulating levels of MMP-9 [11–15] and MMP-10 [21] and TIMP-1 [30–33] in ischemic stroke patients than in controls. In addition, there have been found higher production of TIMP-1 in infarcted brain tissue compared to healthy brain areas [28], and higher expression of TIMP-1 in monocytes of ischemic stroke patients than in healthy controls [29].

In addition, we found higher circulating MMP-10 and TIMP-1 levels, and lower circulating MMP-9 levels in non-surviving severe MMCAI patients than in surviving patients. The findings in respect to MMP-10 are in consonance with a previous study showing an association between serum MMP-10 and functional outcome in

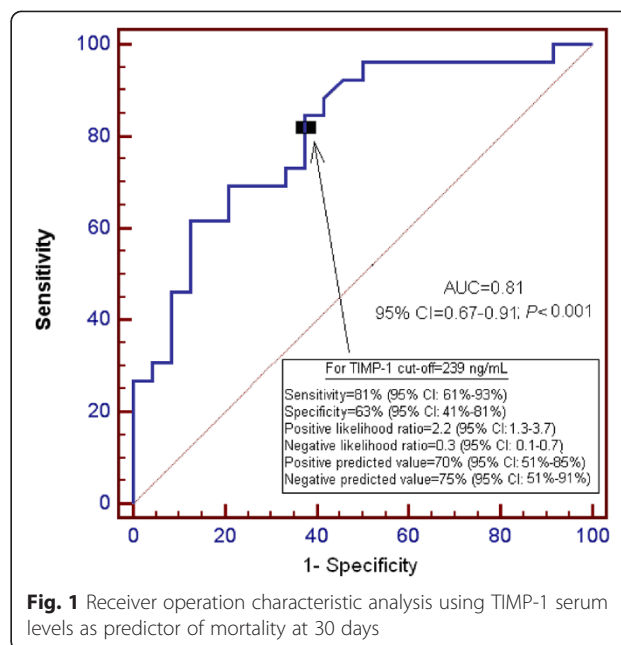


Fig. 1 Receiver operation characteristic analysis using TIMP-1 serum levels as predictor of mortality at 30 days

ischemic stroke patients [21]; however, in our current study, we found for the first time higher serum MMP-10 in non-surviving than in surviving MMCAI patients.

Our findings, showing higher TIMP-1 levels in non-surviving severe MMCAI patients than in surviving patients, could be in agreement with the results of other previous study [20]. A relationship between plasma TIMP-1 levels at 7 days of clinical ischemic with neurological clinical outcome has been demonstrated [20]. Then, another new finding of our study were those higher serum TIMP-1 levels at moment of severe MMCAI diagnosis in non-survivor than in survivor patients. Previously,

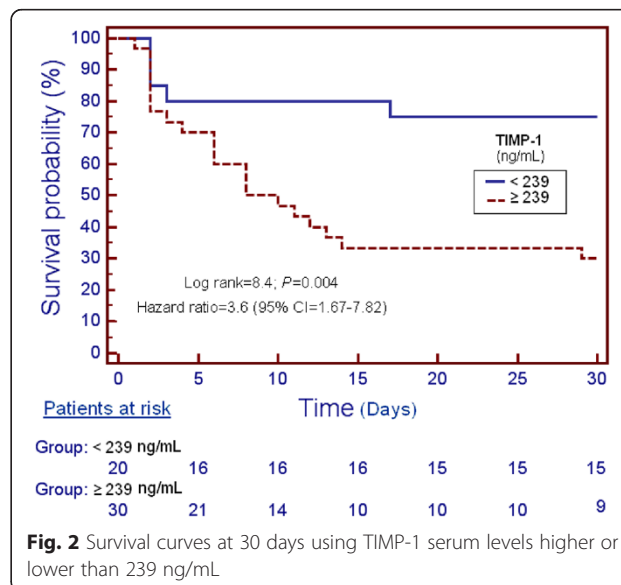


Fig. 2 Survival curves at 30 days using TIMP-1 serum levels higher or lower than 239 ng/mL

Table 3 Multiple binomial logistic regression analysis to predict 30-day mortality

Variable	Odds Ratio	95 % Confidence Interval	P
TIMP-1 > 239 ng/mL	5.82	1.37-24.73	0.02
GCS score	0.79	0.56-1.12	0.19
Age (years)	1.01	0.95-1.07	0.81

there have been found an association between circulating TIMP-1 levels and poor prognosis in elderly men [22], patients with coronary artery disease [23], patients with different types of cancer [24–27], and patients with severe trauma brain injury [34].

On the other hand, the results regarding to circulating MMP-9 levels are in contradiction with those previously published reporting a poor functional outcome with high circulating MMP-9 levels [11–19].

Another interesting new findings of our study were the association between serum TIMP-1 levels and mortality in logistic regression analysis, and the mortality prediction of circulating TIMP-1 levels according to the ROC analysis. These findings agree with the results of a previous study by our team in patients with severe trauma brain injury [34].

The pathophysiological role of circulating TIMP-1 levels in MMCAI patients is still unknown. It is possible that the increased levels in these patients may be due to an increase of circulating MMP-9 and MMP-2 levels, in order to maintain the balance between proteases and inhibitors. Interestingly, we report for the first time an association between circulating levels of TIMP-1 and MMP-10, PAI-1, and TNF-alpha in patients with severe MMCAI patients. Previously higher circulating levels of PAI-1 [37, 38] and TNF-alpha [21] were found in ischemic stroke patients with poor functional outcome. Taken together, these data suggest that TIMP-1 levels could play a role in the pathophysiology of MMCAI. It is possible that increased serum TIMP-1 levels in non-survivors TBI patients is not the cause of death in these patients, rather a biomarker associated with mortality.

Some limitations of our study should be recognized. First, we did not report data about the evolution of TIMPs and MMPs on the time to describe the evolution in non-surviving and surviving TBI patients. Second, the determination of other MMPs and TIMPs would be desirable. Third, the assessment of other inflammatory cytokines and coagulation biomarker could be interesting. Fourth, there is overlap of serum TIMP-1 levels between dead and alive patients at 30 days; thus, the sole use of serum TIMP-1 levels to predict 30-day survival in MMCAI patients should be taken with caution. However, we think that the findings of our study (reporting for the first time an association between TIMP-1 and mortality in MMCAI patients) could open the interest for research about TIMP-1 in MMCAI patients.

The administration of modulators of MMP activity have showed a beneficial effect in rat ischemic stroke models reducing the expression of MMPs, blood–brain barrier leakage, volumen infarction, neurological dysfunction and mortality [39–44]. Thus, from a therapeutic perspective, MMP activity modulators levels could be used as a new class of drugs for the treatment of patients with severe ischemic stroke.

Conclusions

The most relevant and new findings of our study, were that serum TIMP-1 levels in MMCAI patients were associated with mortality, and could be used as a prognostic biomarker of mortality in MMCAI patients.

Abbreviations

MMP: Matrix metalloproteinases; TIMP: Tissue inhibitor of matrix metalloproteinases; ICU: Intensive Care Unit; PaO₂: Pressure of arterial oxygen/fraction inspired oxygen; FIO₂: Pressure of arterial oxygen/fraction inspired oxygen; GCS: Glasgow Coma Scale; INR: International normalized ratio; aPTT: activated partial thromboplastin time; APACHE II: Acute Physiology and Chronic Health Evaluation; PAI: Plasminogen activator inhibitor; TNF: Tumor necrosis factor.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

LL was responsible of conceive, design and coordinate the study, made substantial contributions to acquisition of data, analysis and interpretation of data, and drafted the manuscript. MMM, LR, JJC, JSV, MA, JMBL, JO, JAR, JAP have made substantial contributions to acquisition of data and provided useful suggestions. AJ have made substantial contributions to analysis and interpretation of data. All authors read critically and approved the manuscript, and agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Acknowledgments

This study was supported by fundings from Fundación Canaria de Investigación Sanitaria (FUNCANIS) (La Laguna, Tenerife, Spain) and F.I.M.A. (Pamplona, Navarra, Spain).

Author details

¹Intensive Care Unit, Hospital Universitario de Canarias, Ofra, s/n. La Laguna, 38320 Santa Cruz de Tenerife, Spain. ²Intensive Care Unit, Hospital Universitario Nuestra Señora de Candelaria, Crta del Rosario s/n, Santa Cruz de Tenerife 38010, Spain. ³Intensive Care Unit, Hospital General La Palma, Buenavista de Arriba s/n, Breña Alta, La Palma 38713, Spain. ⁴Intensive Care Unit, Hospital Insular, Plaza Dr. Pasteur s/n, Las Palmas de Gran Canaria 35016, Spain. ⁵Intensive Care Unit, Hospital Universitario Dr. Negrín, Barranco de la Ballena s/n, Las Palmas de Gran Canaria 35010, Spain. ⁶Intensive Care Unit, Hospital Clínico Universitario de Valencia, Avda. Blasco Ibáñez nº17-19, Valencia 46004, Spain. ⁷Research Unit, Hospital Universitario de Canarias, Ofra, s/n. La Laguna, 38320 Santa Cruz de Tenerife, Spain. ⁸Laboratory Department, Hospital Universitario de Canarias, Ofra, s/n. La Laguna, 38320 Santa Cruz de Tenerife, Spain. ⁹Atherosclerosis Research Laboratory, CIMU-University of Navarra, Avda Pío XII nº55, Pamplona 31008, Spain.

Received: 16 February 2015 Accepted: 28 June 2015

Published online: 11 July 2015

References

- Adams Jr HP, del Zoppo G, Alberts MJ, Bhatt DL, Brass L, Furlan A, et al. Guidelines for the early management of adults with ischemic stroke: a guideline from the American Heart Association/American Stroke Association Stroke Council, Clinical Cardiology Council, Cardiovascular Radiology and Intervention Council, and the Atherosclerotic Peripheral Vascular Disease and Quality of Care Outcomes in Research Interdisciplinary Working Groups: the American Academy of Neurology affirms the value of this guideline as an educational tool for neurologists. *Stroke*. 2007;38:1655–711.
- Brinckerhoff CE, Matrisian LM. Matrix metalloproteinases: a tail of a frog that became a prince. *Nat Rev Mol Cell Biol*. 2002;3:207–14.
- Birkedal-Hansen H, Moore WG, Bodden MK, Windsor LJ, Birkedal-Hansen B, DeCarlo A, et al. Matrix metalloproteinases: a review. *Crit Rev Oral Biol Med*. 1993;4:197–250.
- Lorente L, Martín MM, Labarta L, Díaz C, Solé-Violán J, Blanquer J, et al. Matrix metalloproteinase-9, -10, and tissue inhibitor of matrix

- metalloproteinases-1 blood levels as biomarkers of severity and mortality in sepsis. *Crit Care*. 2009;13:R158.
5. Martínez de Lizarrondo S, Roncal C, Calvayrac O, Rodríguez C, Varo N, Purroy A, et al. Synergistic effect of thrombin and CD40 ligand on endothelial matrix metalloproteinase-10 expression and microparticle generation in vitro and in vivo. *Arterioscler Thromb Vasc Biol*. 2012;32:1477–87.
 6. Lorente L, Martín M, Plasencia F, Solé-Violán J, Blanquer J, Labarta L, et al. The 372 T/C genetic polymorphism of TIMP-1 is associated with serum levels of TIMP-1 and survival in patients with severe sepsis. *Crit Care*. 2013;17:R94.
 7. Lorente L, Martín MM, Solé-Violán J, Blanquer J, Labarta L, Díaz C, et al. Association of sepsis-related mortality with early increase of timp-1/mmp-9 ratio. *Plos One*. 2014;9, e94318.
 8. Candelario-Jalil E, Yang Y, Rosenberg GA. Diverse roles of matrix metalloproteinases and tissue inhibitors of metalloproteinases in neuroinflammation and cerebral ischemia. *Neuroscience*. 2009;158:983–94.
 9. Moranchó A, Rosell A, García-Bonilla L, Montaner J. Metalloproteinase and stroke infarct size: role for anti-inflammatory treatment? *Ann N Y Acad Sci*. 2010;1207:123–33.
 10. Ramos-Fernandez M, Bellolio MF, Stead LG. Matrix metalloproteinase-9 as a marker for acute ischemic stroke: a systematic review. *J Stroke Cerebrovasc Dis*. 2011;20:47–54.
 11. Horstmann S, Kalb P, Koziol J, Gardner H, Wagner S. Profiles of matrix metalloproteinases, their inhibitors, and laminin in stroke patients: influence of different therapies. *Stroke*. 2003;34:2165–70.
 12. Lucivero V, Prontera M, Mezzapasa DM, Petruzzellis M, Sancilio M, Tinelli A, et al. Different roles of matrix metalloproteinases-2 and -9 after human ischaemic stroke. *Neurol Sci*. 2007;28:165–70.
 13. Heo JH, Kim YS, Lee KY, Kim EH, Chu CK, Nam JM. Increase in plasma matrix metalloproteinase-9 in acute stroke patients with thrombolysis failure. *Stroke*. 2003;34:e48–50.
 14. Reynolds MA, Kirchik HJ, Dahlen JR, Anderberg JM, McPherson PH, Nakamura KK, et al. Early biomarkers of stroke. *Clin Chem*. 2003;49:1733–9.
 15. Lynch JR, Blessing R, White WD, Grocott HP, Newman MF, Laskowitz DT. Novel diagnostic test for acute stroke. *Stroke*. 2004;35:57–63.
 16. Kim YS, Lee KY, Koh SH, Park CY, Kim HY, Lee YJ, et al. The role of matrix metalloproteinase 9 in early neurological worsening of acute lacunar infarction. *Eur Neurol*. 2006;55:11–5.
 17. Montaner J, Alvarez-Sabín J, Molina C, Anglés A, Abilleira S, Arenillas J, et al. Matrix metalloproteinase expression after human cardioembolic stroke: temporal profile and relation to neurological impairment. *Stroke*. 2001;32:1759–66.
 18. Koh SH, Park CY, Kim MK, Lee KY, Kim J, Chang DJ, et al. Microbleeds and free active MMP-9 are independent risk factors for neurological deterioration in acute lacunar stroke. *Eur J Neurol*. 2011;18:158–64.
 19. Barr TL, Latour LL, Lee KY, Schaeve TJ, Luby M, Chang GS, et al. Blood-brain barrier disruption in humans is independently associated with increased matrix metalloproteinase-9. *Stroke*. 2010;41:e123–8.
 20. Worthmann H, Tryc AB, Goldbecker A, Ma YT, Tountopoulou A, Hahn A, et al. The temporal profile of inflammatory markers and mediators in blood after acute ischemic stroke differs depending on stroke outcome. *Cerebrovasc Dis*. 2010;30:85–92.
 21. Rodríguez JA, Sobrino T, Orbe J, Purroy A, Martínez-Vila E, Castillo J, et al. proMetalloproteinase-10 is associated with brain damage and clinical outcome in acute ischemic stroke. *J Thromb Haemost*. 2013;11:1464–73.
 22. Hansson J, Vasan RS, Ärnlov J, Ingelsson E, Lind L, Larsson A, et al. Biomarkers of extracellular matrix metabolism (MMP-9 and TIMP-1) and risk of stroke, myocardial infarction, and cause-specific mortality: cohort study. *PLoS One*. 2011;6, e16185.
 23. Cavusoglu E, Ruwende C, Chopra V, Yanamadala S, Eng C, Clark LT, et al. Tissue inhibitor of metalloproteinase-1 (TIMP-1) is an independent predictor of all-cause mortality, cardiac mortality, and myocardial infarction. *Am Heart J*. 2006;151:1101.e1–8.
 24. Ylisirnio S, Hoyhtya M, Turpeenniemi-Hujanen T. Serum matrix metalloproteinases -2, -9 and tissue inhibitors of metalloproteinases -1, -2 in lung cancer-TIMP-1 as a prognostic marker. *Anticancer Res*. 2000;20:1311–6.
 25. Wu ZS, Wu Q, Yang JH, Wang HQ, Ding XD, Yang F, et al. Prognostic significance of MMP-9 and TIMP-1 serum and tissue expression in breast cancer. *Int J Cancer*. 2008;122:2050–6.
 26. Yukawa N, Yoshikawa T, Akaike M, Sugimasa Y, Rino Y, Masuda M, et al. Impact of plasma tissue inhibitor of matrix metalloproteinase-1 on long-term survival in patients with colorectal cancer. *Oncology*. 2007;72:205–8.
 27. Yoshikawa T, Cho H, Tsuburaya A, Kobayashi O. Impact of plasma tissue inhibitor of metalloproteinase-1 on long-term survival in patients with gastric cancer. *Gastric Cancer*. 2009;12:31–6.
 28. Cuadrado E, Rosell A, Penalba A, Slevin M, Alvarez-Sabín J, Ortega-Aznar A, et al. Vascular MMP-9/TIMP-2 and neuronal MMP-10 up-regulation in human brain after stroke: a combined laser microdissection and protein array study. *J Proteome Res*. 2009;8:3191–7.
 29. Kouwenhoven M, Carlström C, Ozenci V, Link H. Matrix metalloproteinase and cytokine profiles in monocytes over the course of stroke. *J Clin Immunol*. 2001;21:365–75.
 30. Chen S, Martens-Lobenhoffer J, Weissenborn K, Kielstein JT, Lichtinghagen R, Deb M, et al. Association of dimethylarginines and mediators of inflammation after acute ischemic stroke. *J Neuroinflammation*. 2012;9:251.
 31. Jaroslav P, Christian R, Stefan O, Alexander Z, Zepper P, Holger P, et al. Evaluation of serum biomarkers for patients at increased risk of stroke. *Int J Vasc Med*. 2012;2012:906954.
 32. Pelisek J, Rudelius M, Zepper P, Poppert H, Reeps C, Schuster T, et al. Multiple biological predictors for vulnerable carotid lesions. *Cerebrovasc Dis*. 2009;28:601–10.
 33. Park SY, Kim MH, Kang SY, Suh JT, Lee WI. Inflammatory marker expression and its implication in Korean ischemic stroke patients. *Korean J Lab Med*. 2007;27:197–204.
 34. Lorente L, Martín MM, López P, Ramos L, Blanquer J, Cáceres JJ, et al. Association between serum tissue inhibitor of matrix metalloproteinase-1 levels and mortality in patients with severe brain trauma injury. *PLoS One*. 2014;9, e94370.
 35. Teasdale G, Jennett B. Assessment of coma and impaired consciousness. A practical scale. *Lancet*. 1974;2:81–4.
 36. Knaus WA, Draper EA, Wagner DP, Zimmerman JE. APACHE II: a severity of disease classification system. *Crit Care Med*. 1985;13:818–29.
 37. Vilas D, Gomis M, Blanco M, Cortés J, Millán M, Pérez de la Ossa N, et al. Circadian rhythms in the efficacy of intravenous alteplase in patients with acute ischemic stroke and middle cerebral artery occlusion. *Chronobiol Int*. 2012;29:1383–9.
 38. Raicević R, Jovčić A, Mandić-Radić S, Dordević D, Magdić B, Marković L, et al. Predictive value of changes in the hemostasis system in patients with ischemic brain diseases. *Vojnosanit Pregl*. 2002;59:377–84.
 39. Wang Z, Xue Y, Jiao H, Liu Y, Wang P. Doxycycline-mediated protective effect against focal cerebral ischemia-reperfusion injury through the modulation of tight junctions and PKC δ signaling in rats. *J Mol Neurosci*. 2012;47:89–100.
 40. Leonardo CC, Eakin AK, Ajmo JM, Collier LA, Pennypacker KR, Strongin AY, et al. Delayed administration of a matrix metalloproteinase inhibitor limits progressive brain injury after hypoxia-ischemia in the neonatal rat. *J Neuroinflammation*. 2008;5:34.
 41. Yrjänheikki J, Keinänen R, Pellikka M, Hökfelt T, Koistinaho J. Tetracyclines inhibit microglial activation and are neuroprotective in global brain ischemia. *Proc Natl Acad Sci U S A*. 1998;95:15769–74.
 42. Cai Z, Lin S, Fan LW, Pang Y, Rhodes PG. Minocycline alleviates hypoxic-ischemic injury to developing oligodendrocytes in the neonatal rat brain. *Neuroscience*. 2006;137:425–35.
 43. Fan LW, Lin S, Pang Y, Rhodes PG, Cai Z. Minocycline attenuates hypoxia-ischemia-induced neurological dysfunction and brain injury in the juvenile rat. *Eur J Neurosci*. 2006;24:341–50.
 44. Machado LS, Kozak A, Ergul A, Hess DC, Borlongan CV, Fagan SC. Delayed minocycline inhibits ischemia-activated matrix metalloproteinases 2 and 9 after experimental stroke. *BMC Neurosci*. 2006;7:56.