# **CASE REPORT**



# Autoimmune glial fibrillary acidic protein astrocytopathy associated with breast cancer: a case report

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# Abstract

**Background** Autoimmune glial fibrillary acidic protein (GFAP) astrocytopathy (GFAP-A) is an autoimmune inflammatory central nervous system disorder characterized by the detection of autoantibodies that recognize GFAP in CSF. The pathogenesis of GFAP-A is poorly understood. Some patients had a neoplasm detected and GFAP expressed by neoplasms is plausible as immunogen triggering paraneoplastic neurological autoimmunity.

**Case presentation** We report a case of 76-year-old female patient with GFAP-A complicated with breast cancer. She presented with altered consciousness, nuchal rigidity, speech disturbances, and weakness. Her clinical symptoms were improved by immunotherapy and cancer treatments. Immunohistochemical analysis showed that the restricted tumor expressed GFAP. The infiltration of CD3 +T cells were observed in the peritumoral and intratumoral areas. The most common infiltrating lymphocytes were CD8 +T cells. CD4 +T cells and CD20 + B cells were also observed in the predominant peritumoral area.

**Conclusions** These results suggest that GFAP-A may occur in a paraneoplastic neurological syndrome associated with breast cancer.

Keywords Astrocytopathy, Autoantibody, Breast cancer, Glial fibrillary acidic protein (GFAP)

# Introduction

Autoimmune glial fibrillary acidic protein (GFAP) astrocytopathy (GFAP-A) was first reported as an autoimmune inflammatory central nervous system disorder characterized by the detection of immunoglobulin G (IgG) antibodies that recognize GFAP, which is the main

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intermediate filament protein in mature astrocytes [1].

Further studies confirmed that tissue- and cell-based

testing, using cerebrospinal fluid (CSF), determined the

highest specificity for inflammatory central nervous sys-

tem (CNS) diseases [2, 3]. In some GFAP-A patients,

GFAP-IgG is only detected in CSF, while patients with GFAP-IgG in serum only (not in CSF) have diverse neurological phenotypes that may or may not have an autoimmune cause. The use of one assay alone may yield

nonspecific results. The sensitivity and specificity of the

"GFAP pattern" in tissue-based testing for predicting

GFAP-IgG were previously reported as 95.2% (95% CI

76.2-99.9) and 90.0% (95% CI 55.5-99.8), respectively

[4]. At present, the diagnostic criteria for GFAP-A have

not been established and detection of CSF GFAP-IgG

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by tissue- and cell-based testing is essential for a diagnosis of GFAP-A [2, 3]. In recent years, the clinical features of GFAP-A have become better understood [2, 3, 5-9]. The common phenotype of this disorder includes meningoencephalitis with or without myelitis, and the common prodromal symptoms are fever and headache. During the clinical course, patients present with consciousness disturbances, meningeal irritation, ataxia, involuntary movements such as tremor and myoclonus, urinary dysfunction, cognitive dysfunction, and respiratory failure. Blurred vision related to optic disc edema is also observed [8, 9]. A recent French GFAP-A cohort study found the median age at onset to be 43 years, and that 65% of patients were men. Infectious prodromal symptoms were found in 82% of patients. The most frequent presentation was subacute meningoencephalitis (85%), while cerebellar dysfunction was observed in 57% of cases [9]. There is currently no consensus on treatment regimens for this disorder. The response to corticosteroid therapy is generally good. Sometimes intravenous immunoglobulin therapy or plasma exchange may be performed in addition to corticosteroid therapy. However, some patients experience relapses with bad prognoses, including death. Treatments for refractory or relapsing cases include mycophenolate mofetil, azathioprine, rituximab, cyclophosphamide, and tacrolimus [9, 10].

Cerebrospinal fluid (CSF) can be probed for lymphocyte-predominant pleocytosis and elevated protein levels. Brain magnetic resonance imaging (MRI) can show abnormal hyperintensity lesions on T2-weighted and fluid-attenuated inversion recovery (FLAIR) images [8]. Brain linear perivascular radial gadolinium-enhancement (LPRGE) patterns, an imaging hallmark of GFAP-A, are observed in about half of all patients [2, 8].

The pathogenesis of GFAP-A is poorly understood. Pathologically, there is a marked lymphocytic infiltration of the brain parenchyma, with many CD8+and CD4+T cells, especially in the perivascular area [11–13]. GFAPspecific cytotoxic T cells are also likely effectors of this disorder [2, 14]. Approximately 20% of patients have a neoplasm [14]. GFAP expressed by neoplasia is plausible as an immunogen-triggering paraneoplastic neurological autoimmunity. The most commonly detected neoplasia is ovarian teratoma [2]. Other neoplasms are rare and diverse. Herein, we report the clinical features and the pathological findings of tumor tissue in a GFAP-A patient with breast cancer.

# **Case report**

A 76-year-old woman who had a past medical history of hypertension, atrial fibrillation, and right thalamic hemorrhage was admitted to the hospital because of fever and impaired balance. She suffered neck pain and dysarthria two days after admission and became somnolent. CSF examination showed pleocytosis (130 cells/mm<sup>3</sup> [mononuclear cell: 130 cells/mm<sup>3</sup>], normal <5 cells/mm<sup>3</sup>) and increased protein levels (113 mg/dL, normal<50 mg/ dL). She was treated with intravenous acyclovir on suspicion of viral meningoencephalitis. Her symptoms did not improve, and she was referred to our hospital for further investigation and treatment 22 days after onset. On admission, she was afebrile, and her consciousness level was E3V3M5 on the Glasgow Coma Scale. Her speech was slurred and barely comprehensible. She had left hemiparesis because of a past thalamic hemorrhage and flaccid muscle weakness in her right lower extremity. Tendon reflexes were hyperreflexia in both upper limbs and areflexia in both lower limbs. She had nuchal rigidity, but Kernig's sign and Brudzinski's sign were not observed.

Peripheral blood cell counts showed mild thrombocytopenia ( $141 \times 10^3 / \mu$ L, normal range:  $158 - 348 \times 10^3 / \mu$ L). Biochemical examinations showed hypoalbuminemia (2.8 g/dL, normal range: 4.1-5.1 g/dL) and an elevated urea-nitrogen creatinine ratio suggestive of dehydration. C-reactive protein was within the normal limit. Serum thyroid stimulating hormone and free thyroxine levels were within normal limits. A mild elevated carbohydrate antigen 19-9 level was observed (39.2 U/mL, normal≤37.0 U/mL). Serum anti-nuclear antibody and anti-aquaporin 4 antibody were negative. Anti-neuronal antibodies including anti-amphiphysin, CV2, Ma2, Ri, Yo, Hu, recoverin, SRY-related HMG-box gene 1, titin, zinc-finger protein of the cerebellum 4, Tr, and glutamic acid decarboxylase 65 antibodies were all negative results (Euroimmun, Lübeck, Germany). Mycobacterium tuberculosis specific interferon-gamma release assay and serum Candida, Aspergillus, and Cryptococcus antigens were negative.

CSF examination showed normal opening pressure (105 mmH<sub>2</sub>O), pleocytosis (42 cells/mm<sup>3</sup> [mononuclear cell: 41 cells/mm<sup>3</sup>]), increased protein levels (95 mg/dL), and mildly decreased glucose levels (40 mg/dL). Bacterial culture had a negative result. Herpes simplex virus and *Mycobacterium tuberculosis* polymerase chain reaction tests also gave negative results. CSF cytology showed no malignant cells. Later, CSF GFAP-IgG was detected by transfected cell-based assay and tissue-based immuno-fluorescence assay according to previous reports [2, 5] (Fig. 1).

Brain MRI scans showed abnormal signal changes caused by a past right thalamic hemorrhage (Fig. 2A) and extended white matter hyperintensity lesions in the deep and periventricular white matter in the right frontal and parietal lobes on T2-weighted and FLAIR images (Fig. 2B). Gadolinium contrast-enhanced brain MRI scans showed heterogeneous thickening of the dura mater (Fig. 2C). Spinal MRI showed no abnormal



Fig. 1 Detection of cerebrospinal fluid (CSF) glial fibrillary acidic protein (GFAP) immunoglobulin G (IgG).

Cell-based assay of GFAPa-transfected HEK293 cells (AC). GFAP-IgG was detected in the CSF of the patient with autoimmune GFAP astrocytopathy (A). HEK293 cells stably express green fluorescent protein (GFP)-tagged GFAPa (B). Colocalization of the patient's CSF-IgG and GFAPa is yellow in merged images (C)

Tissue-based immunofluorescence assay (D–F). Immunoreactivity of the patient's CSF-IgG was observed in astrocytes of the pial, subpial (D), parenchyma (E), and cerebellum (F)

signal changes in the spinal cord. Whole body computed tomography (CT) showed no findings of neoplasia. However, early-stage breast cancer was found in the left breast on mammography.

Based on these results, the patient was diagnosed with GFAP-A. She was treated with an intravenous infusion of 1 gram per day methylprednisolone for 3 days starting on day 8 after admission. Her nuchal rigidity disappeared on day 10. Her consciousness level gradually improved starting on day 11 and became completely clear on day 32. She was also treated with intravenous immunoglobulin (0.4 gram per kilogram body weight for 5 days) on day 36 and again on day 60. She was temporarily transferred to the local hospital on day 91 for rehabilitation.

A month later, the patient was re-admitted to our hospital and underwent a simple mastectomy for breast cancer. The pathological findings were invasive ductal carcinoma, tubule forming type (Fig. 3A, B). Immunohistochemical analysis showed that the restricted tumor expressed GFAP (Fig. 3C, D). The infiltration of CD3+T cells were observed in the peritumoral and intratumoral areas (Fig. 3E). The most common infiltrating lymphocytes were CD8+T cells (Fig. 3F). CD4+T cells and CD20+B cells were also observed in the predominant peritumoral area (Fig. 3G, H). Her condition did not deteriorate, and no relapse occurred thereafter.

## Discussion

Here, we provide a report on a GFAP-A patient with breast cancer. This patient was initially suspected to have viral meningoencephalitis and treated with an antiviral therapy, but her symptoms did not improve. Afterwards, she was diagnosed with GFAP-A, and her condition



Fig. 2 Brain magnetic resonance imaging

Brain magnetic resonance T2-weighted images show abnormal signal changes caused by a past right thalamic hemorrhage (arrow) (A) and extended white matter hyperintensity lesions in the deep and periventricular white matter in the right frontal and parietal lobes (B). Gadolinium contrast-enhanced brain MRI scans show heterogeneous thickening of the dura mater (arrows) (C)

improved after receiving immunotherapy and cancer treatments.

Previously, Flanagan et al. reported that diverse neoplasms were detected in 22 of 102 patients subsequent to neurological presentation [2]. The most commonly detected neoplasia in these cases was ovarian teratoma. Another previous report described that an ovarian teratoma from a patient with GFAP-A with coexisting N-Methyl-D-Aspartate Receptor (NMDAR)-IgG had robust GFAP staining and sparse NMDAR staining [5]. Another group also reported that an ovarian teratoma associated with coexisting NMDAR and GFAP autoimmune meningoencephalitis in an adolescent girl had extensive CD3+T cell infiltration [16]. In contrast, many other types of neoplasms have been reported in this condition, including adenocarcinoma (endometrium, esophagus, kidney, prostate, colon, ovary, lung, breast), gliomas, multiple myeloma, pleomorphic parotid adenoma, carcinoid, squamous cell carcinoma (head, neck, nasopharyngeal), small cell lung carcinoma, Hodgkin lymphoma, B cell lymphoma, chronic lymphocytic leukemia, melanoma, renal cell carcinoma, breast ductal carcinoma, urothelial bladder carcinoma, meningioma, and thymoma [2-6, 17]. The true association between these neoplasms and GFAP-A has not been clarified. However, in this study, we have provided the first identification of GFAP expression in breast cancer tumor tissue and the extensive infiltration of CD3+T cells, including CD8+T cells, CD4+T cells, and CD20+B cells in peritumoral and intratumoral areas. Although we have not confirmed that GFAP is absent in breast cancer cells without GFAP-A, database searches (https://www.proteinatlas.org) show that GFAP is not expressed in breast cancer cell lines [18]. We suggest that the breast cancer is an immunogen that triggers GFAP-A. When GFAP-A is diagnosed in adult females, mammary glands should be included in screening for neoplasms. In addition, meningoencephalitis, with or without myelitis, in patients with breast cancer may be due to this syndrome, and therefore, the CSF levels of GFAP-IgG should be examined. Although the pathophysiological mechanisms of GFAP-A remain to be elucidated, GFAP-specific CD8+T cells are likely effectors of this disorder [2]. It is possible that ectopic expression of GFAP in the neoplasm triggered the autoimmune response against GFAP including the production of GFAP-specific CD8+T cells and B cells. After disruption of the blood-brain barrier by infection, these cells can infiltrate the CNS and cause GFAP-specific CD8+T cellrelated inflammation and GFAP-IgG production.

In the patient described in this study, gadolinium contrast-enhanced brain MRI scans showed a heterogeneous thickening of the dura mater. Similarly, a recent report



#### Fig. 3 Breast cancer pathology

Hematoxylin-Eosin (HE) staining (A) and immunohistochemical analysis using commercial antibodies against pan-cytokeratin AE1/AE3 (B) show invasive ductal carcinoma. Immunohistochemical analysis using commercial antibodies against GFAP (C) and the merged image of staining AE1/AE3 and GFAP (D) reveals the expression of GFAP in breast tumor tissue. Immunohistochemical analysis shows that the infiltration of CD3 +T cells were observed in peritumoral and intratumoral areas (E). The most common infiltrating lymphocytes were CD8 +T cells (F). CD4 +T cells (G) and CD20 + B cells (H) were also observed in predominant peritumoral areas

described a pediatric patient with hypertrophic pachymeningitis associated with GFAP-A, and the authors indicate that hypertrophic pachymeningitis may be one of the clinical phenotypes for GFAP-A [19], which is also corroborated by our findings.

The limitations of this study are as follows. First, this study is a case report and lacks information on other potential risk factors for GFAP-A. To further examine the association between breast cancer and GFAP-A, it will be necessary to identify additional GFAP-A patients who also present with breast cancer. Second, differences in inflammatory cells infiltrating tumor tissues of breast cancers with and without GFAP-A are not known. Differences between these tumor tissues and the presence of GFAP antigen-specific lymphocytes in tumor tissue with GFAP-A warrant further investigation.

# Conclusion

Here, we report a patient with GFAP astrocytopathy, which may occur in a paraneoplastic neurological syndrome associated with breast cancer.

#### Abbreviations

CSF	cerebrospinal fluid
СТ	computed tomography
FLAIR	fluid-attenuated inversion recovery

gfap gfap-a Igg Lprge Mri	glial fibrillary acidic protein Autoimmune glial fibrillary acidic protein astrocytopathy immunoglobulin G linear perivascular radial gadolinium-enhancement magnetic resonance imaging
MRI	magnetic resonance imaging
NMDAR	N-Methyl-D-Aspartate Receptor

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#### Author contributions

TY and AK designed the study, collected the data, and drafted the manuscript. AT, MM, and HT analyzed the data. TS supervised this study. All authors read and approved the final manuscript.

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#### **Data Availability**

All data reported within the article are available as an anonymized set by request from qualified investigators.

### Declarations

#### Ethical approval and consent to participate

We examined CSF GFAP-IgG under a study approved by the institutional review board of Gifu University Graduate School of Medicine, Gifu, Japan (27–43). Informed consent was obtained from the patient for the use of CSF samples and for the publication of the case report.

#### **Consent for publication**

All the authors have read the manuscript entitled "Autoimmune glial fibrillary acidic protein astrocytopathy associated with breast cancer" and have approved this submission. The informed consent of the patient of this study to publish her identifiable data in an online, open-access journal was obtained.

#### **Competing interests**

The authors declare no competing interests.

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#### References

- Fang B, McKeon A, Hinson SR, Kryzer TJ, Pittock SJ, Aksamit AJ, Lennon VA. Autoimmune glial fibrillary acidic protein astrocytopathy: a novel Meningoencephalomyelitis. JAMA Neurol. 2016;73:1297–307.
- Flanagan EP, Hinson SR, Lennon VA, Fang B, Aksamit AJ, Morris PP, Basal E, Honorat JA, Alfugham NB, Linnoila JJ, Weinshenker BG, Pittock SJ, McKeon A. Glial fibrillary acidic protein immunoglobulin G as biomarker of autoimmune astrocytopathy: analysis of 102 patients. Ann Neurol. 2017;81:298–309.
- Kunchok A, Zekeridou A, McKeon A. Autoimmune glial fibrillary acidic protein astrocytopathy. Curr Opin Neurol. 2019;32:452–8.
- Nagata N, Kanazawa N, Mitsuhata T, Iizuka M, Nagashima M, Nakamura M, Kaneko J, Kitamura E, Nishiyama K, Iizuka T. Neuronal surface antigen-specific immunostaining pattern on a rat brain immunohistochemistry in autoimmune encephalitis. Front Immunol. 2023;13:1066830.
- Dubey D, Hinson SR, Jolliffe EA, Zekeridou A, Flanagan EP, Pittock SJ, Basal E, Drubach DA, Lachance DH, Lennon VA, McKeon A. Autoimmune GFAP astrocytopathy: prospective evaluation of 90 patients in 1 year. J Neuroimmunol. 2018;321:157–63.
- Iorio R, Damato V, Evoli A, Gessi M, Gaudino S, Di Lazzaro V, Spagni G, Sluijs JA, Hol EM. Clinical and immunological characteristics of the spectrum of GFAP autoimmunity: a case series of 22 patients. J Neurol Neurosurg Psychiatry. 2018;89:138–46.
- Kimura A, Takekoshi A, Yoshikura N, Hayashi Y, Shimohata T. Clinical characteristics of autoimmune GFAP astrocytopathy. J Neuroimmunol. 2019;332:91–8.
- Yang X, Liang J, Huang Q, Xu H, Gao C, Long Y, Xiao X. Treatment of autoimmune glial fibrillary acidic protein astrocytopathy: follow-up in 7 cases. Neuroimmunomodulation. 2017;24:113–9.
- Dumonceau AG, Ameli R, Rogemond V, Ruiz A, Joubert B, Muñiz-Castrillo S, Vogrig A, Picard G, Ambati A, Benaiteau M, Rulquin F, Ciron J, Deiva K, de Broucker T, Kremer L, Kerschen P, Sellal F, Bouldoires B, Genet R, Biberon J,

- 10. Xiao J, Chen X, Shang K, Tang Y, Chen M, Deng G, Qin C, Tian DS. Clinical, neuroradiological, diagnostic and prognostic profile of autoimmune glial fibrillary acidic protein astrocytopathy: a pooled analysis of 324 cases from published data and a single-center retrospective study. J Neuroimmunol. 2021;360:577718.
- Long Y, Liang J, Xu H, Huang Q, Yang J, Gao C, Qiu W, Lin S, Chen X. Autoimmune glial fibrillary acidic protein astrocytopathy in chinese patients: a retrospective study. Eur J Neurol. 2018;25:477–83.
- Shu Y, Long Y, Chang Y, Li R, Sun X, Wang Y, Huang Y, Li J, Chen J, Yang Y, Lu Z, Hu X, Kermode AG, Qiu W. Brain immunohistopathology in a patient with autoimmune glial fibrillary acidic protein astrocytopathy. Neuroimmunomodulation. 2018;25:1–6.
- Yuan Z, Li H, Huang L, Fu C, Chen Y, Zhi C, Qiu W, Long Y. CD8 + T-cell predominance in autoimmune glial fibrillary acidic protein astrocytopathy. Eur J Neurol. 2021;28:2121–5.
- 14. Zekeridou A, McKeon A, Flanagan EP. A path to understanding autoimmune GFAP astrocytopathy. Eur J Neurol. 2018;25:421–2.
- Graus F, Vogrig A, Muñiz-Castrillo S, Antoine JG, Desestret V, Dubey D, Giometto B, Irani SR, Joubert B, Leypoldt F, McKeon A, Prüss H, Psimaras D, Thomas L, Titulaer MJ, Vedeler CA, Verschuuren JJ, Dalmau J, Honnorat J. Updated diagnostic criteria for paraneoplastic neurologic syndromes. Neurol Neuroimmunol Neuroinflamm. 2021;8:e1014.
- Martin AL, Jolliffe E, Hertweck SP. Ovarian Teratoma Associated with Coexisting Anti-N-Methyl-D-Aspartate receptor and glial fibrillary acidic protein autoimmune meningoencephalitis in an adolescent girl: a Case Report. J Pediatr Adolesc Gynecol. 2018;31:321–4.
- 17. Huang Q, Yang H, Liu T, Xu H, Chen B, Liu S, Li W, Long Y, Gao C. Patients with suspected benign tumors and glial fibrillary acidic protein autoantibody: an analysis of five cases. Int J Neurosci. 2019;129:1183–8.
- Pontén F, Jirström K, Uhlen M. The human protein Atlas–a tool for pathology. J Pathol. 2008;216:387–93.
- Tan C, Zhong M, Yao Z, Hong S, Jiang L, Jiang Y. Anti-GFAP Antibody-Associated Hypertrophic Pachymeningitis. Neuropediatrics. 2022;53:143–5.

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