

STUDY PROTOCOL

Open Access



Natural history, outcome measures and trial readiness in LAMA2-related muscular dystrophy and SELENON-related myopathy in children and adults: protocol of the LAST STRONG study

Karlijn Bouman^{1,2*} , Jan T. Groothuis³, Jonne Doorduyn¹, Nens van Alfen¹, Floris E. A. Udink ten Cate⁴, Frederik M. A. van den Heuvel⁵, Robin Nijveldt⁵, Willem C. M. van Tilburg⁶, Stan C. F. M. Buckens⁶, Anne T. M. Dittrich⁷, Jos M. T. Draaisma⁷, Mirian C. H. Janssen⁸, Erik-Jan Kamsteeg⁹, Esmee S. B. van Kleef¹, Saskia Koene¹⁰, Jan A. M. Smeitink¹¹, Benno Küsters¹², Florence H. J. van Tienen¹³, Hubert J. M. Smeets^{13,14,15}, Baziel G. M. van Engelen¹, Corrie E. Erasmus^{2†} and Nicol C. Voermans^{1†}

Abstract

Background: SELENON (SEPN1)-related myopathy (SELENON-RM) is a rare congenital myopathy characterized by slowly progressive proximal muscle weakness, early onset spine rigidity and respiratory insufficiency. A muscular dystrophy caused by mutations in the *LAMA2* gene (LAMA2-related muscular dystrophy, LAMA2-MD) has a similar clinical phenotype, with either a severe, early-onset due to complete Laminin subunit $\alpha 2$ deficiency (merosin-deficient congenital muscular dystrophy type 1A (MDC1A)), or a mild, childhood- or adult-onset due to partial Laminin subunit $\alpha 2$ deficiency. For both muscle diseases, no curative treatment options exist, yet promising preclinical studies are ongoing. Currently, there is a paucity on natural history data and appropriate clinical and functional outcome measures are needed to reach trial readiness.

* Correspondence: Karlijn.bouman@radboudumc.nl

†Corrie E. Erasmus and Nicol C. Voermans contributed equally to this work.

¹Department of Neurology, Donders Institute for Brain, Cognition and Behaviour, Radboud university medical center, Nijmegen, The Netherlands

²Department of Pediatric Neurology, Donders Institute for Brain, Cognition and Behaviour, Amalia Children's Hospital, Radboud university medical center, Nijmegen, The Netherlands

Full list of author information is available at the end of the article



© The Author(s). 2021 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Methods: LAST STRONG is a natural history study in Dutch-speaking patients of all ages diagnosed with SELENON-RM or LAMA2-MD, starting August 2020. Patients have four visits at our hospital over a period of 1.5 year. At all visits, they undergo standardized neurological examination, hand-held dynamometry (age ≥ 5 years), functional measurements, questionnaires (patient report and/or parent proxy; age ≥ 2 years), muscle ultrasound including diaphragm, pulmonary function tests (spirometry, maximal inspiratory and expiratory pressure, sniff nasal inspiratory pressure; age ≥ 5 years), and accelerometry for 8 days (age ≥ 2 years); at visit one and three, they undergo cardiac evaluation (electrocardiogram, echocardiography; age ≥ 2 years), spine X-ray (age ≥ 2 years), dual-energy X-ray absorptiometry (DEXA)-scan (age ≥ 2 years) and full body magnetic resonance imaging (MRI) (age ≥ 10 years). All examinations are adapted to the patient's age and functional abilities. Correlation between key parameters within and between subsequent visits will be assessed.

Discussion: Our study will describe the natural history of patients diagnosed with SELENON-RM or LAMA2-MD, enabling us to select relevant clinical and functional outcome measures for reaching clinical trial-readiness. Moreover, our detailed description (deep phenotyping) of the clinical features will optimize clinical management and will establish a well-characterized baseline cohort for prospective follow-up.

Conclusion: Our natural history study is an essential step for reaching trial readiness in SELENON-RM and LAMA2-MD.

Trial registration: This study has been approved by medical ethical reviewing committee Region Arnhem-Nijmegen (NL64269.091.17, 2017–3911) and is registered at [ClinicalTrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT04478981) (NCT04478981).

Keywords: LAMA2, Laminin subunit $\alpha 2$ deficiency, Merosin-deficient congenital muscular dystrophy type 1A (MDC1A), SELENON, SEPN1, Natural history, Outcome measures, Trial readiness, All ages

Background

Selenoprotein N-related congenital myopathy (SEPN1 or SELENON-RM) is a rare congenital myopathy with an estimated prevalence of 0.5 in 1000,000 [1]. Core features include slowly progressive axial muscle weakness, early-onset rigidity of the spine, scoliosis and respiratory insufficiency. Delayed motor development is the most common presenting sign. Muscle biopsies show multinicores as the most common lesion, often associated with mild dystrophic features [2]. Laminin $\alpha 2$ -related muscular dystrophy (LAMA2-MD) has a similar clinical phenotype, with an estimated prevalence of 4 in 500,000 [3]. It has a heterogeneous disease spectrum ranging from a severe, early-onset congenital muscular dystrophy (complete Laminin subunit $\alpha 2$ deficiency, also called merosin-deficient congenital muscular dystrophy type 1A (MDC1A)) to a mild, childhood- or adult-onset limb-girdle type muscular dystrophy (partial Laminin subunit $\alpha 2$ deficiency). Additionally, patients may suffer from epileptic seizures and may show characteristic diffuse brain white matter lesions on magnetic resonance imaging (MRI) [4]. The clinical diagnosis of SELENON-RM and LAMA2-MD is confirmed by recessive pathogenic variants in the *SELENON* gene (OMIM-number 606210) or *LAMA2* gene (OMIM-number 156225), respectively [5–9]. Currently, no curative treatment options exist for neither SELENON-RM nor LAMA2-MD. Optimal pulmonary, cardiac, nutrition and orthopedic management, in combination with supportive care from rehabilitation and allied health care, is essential for

preventing or treating severe complications [10, 11]. Further, promising new therapies are currently being developed [12–21].

Selenoprotein N is an endoplasmic reticulum (ER) calcium sensor that responds to diminished luminal calcium levels by refilling the ER calcium stores [22]. SELENON-RM has striking similarities at the cellular level with classical mitochondrial diseases. This has led to the hypothesis that sonlicromanol, a new clinical stage chemical entity with a dual activity as antioxidant and redox modulator developed for mitochondrial oxidative phosphorylation disturbances, is also beneficial for patients with SELENON-RM [12, 13]. Interestingly, the first results of experiments in an animal model (SELENON knock-out zebrafish) showed improved muscular function (unpublished data). Further, other antioxidants are also hypothesized to be beneficial in the treatment of patients with SELENON-RM [13–15]. In general, the recognition of the interplay between mitochondrial bioenergetics and endoplasmic reticulum paves the way to the identification of potential treatment options [16, 17]. Laminin subunit $\alpha 2$ is an extracellular matrix protein that links with dystrophin on the inner side of the muscle membrane. This linkage is of high importance for normal skeletal muscle function as it stabilizes the sarcolemma and protects the muscle fibers from contraction-induced damage [23, 24]. Further, a metabolic impairment, with reduced mitochondrial respiration and enhanced glycolysis, was observed in human Laminin subunit $\alpha 2$ deficient muscle cells [25]. For

LAMA2-MD, preclinical studies on the use of linker proteins, on exogenous administration of Laminin-111, on upregulation of LAMA1, on genome editing technology and on the use of antioxidant molecules are being performed in animal models [18–21, 26, 27]. Moreover, different research groups are working on revealing the precise pathophysiology, which could eventually help to design new treatment strategies for SELENON-RM [14, 28–31] or LAMA2-MD [32–34].

In order to pave the way towards clinical trials, it is essential to identify and characterize the patients clinically and genetically, and to select clinical and functional outcome measures that correlate with muscle function and that are sensitive to change over time. These outcome measures can be used to determine the effectivity of possible treatment options in clinical trials. Three large clinical studies have recently been performed, one in SELENON-RM and two in LAMA2-MD. We discuss the main findings below.

In SELENON-RM patients, a retrospective clinical, histologic and genetic analysis of 132 pediatric and adult patients (age range at last examination 2 to 58 years) was performed by Villar-Quiles et al. [2]. The main prognostic determinants for disease severity included scoliosis and respiratory management, body mass abnormalities and the specific *SELENON* mutation found in the patient. The latter indicates a genotype-phenotype association between bi-allelic null mutations and more severe disease. This study reports the largest SELENON-RM series and the first one including pediatric, adolescent and adult patients followed-up for several decades. Limitations of this study are its retrospective design, and the absence of functional measurements and convenient muscle visualizing techniques (i.e. muscle ultrasound or MRI) performed in a standardized manner.

In LAMA2-MD patients, a 5-year prospective natural history study that included 24 patients (age range 4 to 22 years) was performed by Jain et al. [35, 36]. The MFM-32 was found to be sensitive to change in ambulatory and non-ambulatory patients with LAMA2-MD. In non-ambulatory patients, they found a yearly decline in knee flexion strength and passive range of motion (PROM) of left elbow extension. Limitations of this study included the small subset of examinations performed and the absence of convenient muscle visualizing techniques. Further, patients were selected based on a convenience sample, possibly leading to a selection bias. Moreover, the age range limits the availability on natural history data in the very young children (< 4 years) and in older adult patients (> 22 years). Recently, Zambon et al. published a retrospective longitudinal study on 46 patients with LAMA2-MD of whom 42 patients had a complete and 4 patients had a partial Laminin subunit $\alpha 2$ deficiency (age range at last examination 12 to 22

years) [37]. They found a linear decrease in passive range of motion of left elbow extension and a linear decline in percentage predicted forced vital capacity. The intrinsic limitations included the retrospective nature of data collection, the limited age range and the inconsistencies in the use of functional scales throughout follow-up (i.e. frequency of examinations, indication for ancillary examinations etcetera).

In short, a prospective natural history study in an unselected group of patients including a plethora of clinical and functional outcome measures is lacking in both SELENON-RM and LAMA2-MD. Due to the promising ongoing preclinical studies, there is a high need to obtain natural history data in order to reach trial readiness for both muscle diseases. The similarities in the clinical phenotype of both muscle diseases allows us to combine both studies in one study protocol.

Objectives

The primary objectives of this study in order to reach trials readiness, are:

1. to assess 1.5-year natural history in patients with SELENON-RM or LAMA2-MD;
2. to select relevant and sensitive clinical and functional outcome measures.

The secondary objectives of this study are:

3. to provide prevalence estimations of SELENON-RM and LAMA2-MD in the Netherlands and Flanders (Dutch-speaking part of Belgium);
4. to establish a well-characterized baseline cohort of patients with SELENON-RM or LAMA2-MD for prospective follow-up and recruitment for future clinical trials;
5. to assess the clinical features to optimize clinical management for patients with SELENON-RM or LAMA2-MD.

Methods / design

Study design

Our study on LAMA2-MD and SELENON-RM To Study Trial Readiness, Outcome measures and Natural history (LAST STRONG) is a prospective, single-center, observational study with repeated measurements performed at the Department of Neurology and Pediatric Neurology within the neuromuscular center of the Radboud university medical center, The Netherlands. Our center is a tertiary referral center for neuromuscular diseases. Participation in the study will not affect the usual care provided by the patient's own medical team. Patients are invited to visit our hospital four times over a period of 1.5 year, with an interval of six months. During these visits, a predefined subset of investigations will be

performed (See Table 1). Further, medical records from routine clinical care will be requested. Our study has been approved by the medical ethical reviewing committee Region Arnhem – Nijmegen (NL64269.091.17; 2017–3911) and is registered at [ClinicalTrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT04478981) (NCT04478981).

Study population

Based on the prevalence reported in previous studies on SELENON-RM and LAMA2-MD, we expect to identify between 20–30 patients in each disease group. Based on our experience in rare diseases, we estimate that around 50% of the patients will participate in the study. We therefore aim to include 10–15 participants in each disease group. Inclusion criteria include a genetic confirmation of SELENON-RM or LAMA2-MD by two recessive pathologic mutations in the *SELENON* or *LAMA2* gene, respectively, or typical clinical and histological alterations combined with genetic confirmation in a first degree relative. Additionally, patients must be willing and able to complete (part of) the measurement protocol in the Radboud university medical center. If patients do not wish or are not able to visit our center, they are offered to participate in this study by sharing medical records, completing questionnaires, and undergoing a medical history and physical examination through home visits, video and/or telephone interview. Exclusion criteria are an insufficient understanding of the Dutch language and the unwillingness of the patient or his/her legal representatives to provide written informed consent for participation in our study.

Recruitment

Participants will be recruited non-selectively and consecutively in the periods from August 2020 to August 2021 (See Fig. 1). In order to estimate the prevalence and to provide data on the complete spectrum of patients with SELENON-RM or LAMA2-MD, we aim to reach all patients in The Netherlands and the Dutch-speaking part of Belgium (Flanders) in our study. All Dutch and Dutch-speaking Belgian (pediatric) neurologists, rehabilitation specialists and clinical geneticists are personally asked about potential participants. Additionally, all patients known at our own neuromuscular center will be personally informed. Moreover, we will directly recruit participants through promotion of our study on patient information days, social media and through patient organizations.

Demographics

Date of birth, sex, weight (kg), height (m), comorbidity and medication will be recorded.

Genetics

Upon inclusion in our study, all participants (or a first degree relative) have undergone genetic examination as

part of regular diagnostic work-up. The genetics reports, including information on the specific genetic alterations, will be requested. Regarding LAMA2-MD, we expect to mostly include patients harboring the Dutch founder mutation in the *LAMA2* gene (c.5562 + 5G > C). All genetic laboratories in The Netherlands will be contacted to inventory the number of patients diagnosed with SELENON-RM or LAMA2-MD in order to contrast this number against the number of participants in our natural history study.

Muscle biopsy

Pathological records and stained slides will be requested from participants diagnosed with LAMA2-MD in whom muscle biopsy material was previously taken as part of regular diagnostic work-up. Hereby we aim to classify LAMA2-MD patients as either suffering from a complete or a partial Laminin subunit $\alpha 2$ deficiency.

Neurological examination and functional measurements

All patients undergo a standard neurological examination by one assessor (KB). Additionally, muscle strength, facial muscle weakness, reflexes, muscle tone, and dysmorphic features are assessed by two independent assessors (KB and CE or NV). Muscle strength (MRC grading scale) will be assessed of the following muscles: neck flexor, neck extensor, sternocleidomastoid, trapezius, deltoid, biceps brachii, triceps brachii, wrist extensor, wrist flexor, finger extensor, finger flexor, finger spreader, iliopsoas, gluteus maximus, quadriceps, hamstrings, foot dorsiflexor, foot plantarflexor, extensor hallucis longus and toe flexor muscles. Further, muscle strength of the following muscles will be measured using a hand-held dynamometer (Citec, CT3002) [38–41]:

1. Neck flexors and neck extensors: sitting upright; head up at 90° from horizontal
2. Elbow flexors and extensors: supine; shoulder adducted, elbow 90° flexed, forearm supinated
3. Knee extensors: sitting upright; knee 90° flexed
4. Foot plantar- and dorsiflexors: supine; foot 90° dorsiflexed
5. Pinch grip: sitting upright; shoulder adducted, elbow 90° flexed, forearm pronated

Additionally, the passive range of motion (PROM) of the elbow, wrist, hip, knee and ankle joints is assessed by a goniometer [42]. Functional measurements include:

1. The Children's Hospital of Philadelphia Infant Test of Neuromuscular Disorders (CHOP INTEND) (age < 2 years) [43, 44].
 - a. CHOP INTEND has been shown to be valid for the assessment of motor skills of children below 2 years of age.

Table 1 Examinations performed in LAST STRONG study

Outcome domain	Examination	Age	Visit
Medical history			
Past	Perinatal period, motor milestones	All	1
Current	Functional abilities, comorbidities, devices, treatments	All	All
Neurological examination and functional measurements			
Muscle function	Muscle strength assessment (Medical Research Council, MRC)	≥ 5 years	All
	Hand-Held dynamometry (HHD)	≥ 5 years	All
	The Children's Hospital of Philadelphia Infant Test of Neuromuscular Disorders (CHOP INTEND)	< 2 years	All
	Hammersmith Infant Neurological Examinations (HINE)	< 2 years	All
	Motor Function Measurement (MFM)-20/320	≥ 2 years	All
	Hammersmith Functional Motor Scale (HFMS); non-ambulant participants only	≥ 2 years	All
	Pediatric Balance Scale (PBS); ambulant participants only	2–15 years	All
	Mini Balance Evaluation Systems Test (miniBEST); ambulant participants only	≥ 16 years	All
	Graded and timed function tests; ambulant participants only		All
	1. 30 s sit to stand, climb 4 stairs, rise from the floor, timed up and go (TUG)	≥ 2 years	
	2. 6-Minute Walk test (6MWT), 10-Meter Walk test (10MWT)	≥ 5 years	
	Functional Ambulation Classification (FAC)	≥ 5 years	All
	Vignos and Brooke scale	≥ 2 years	All
Contractures	Goniometry	≥ 2 years	All
Other	Coordination, gait, reflexes, cranial nerves and facial muscles, dysmorphic features	≥ 2 years	All
Questionnaires			
Quality of Life	Pediatric Quality of Life Inventory (PedsQL) (Generic Core Scale, Neuromuscular Module; Child Self-report and/or parent proxy)	2–17 years	All
	Research and Development-36 (RAND36)	≥ 18 years	All
	Individualized Neuromuscular Quality of Life (INQoL)	≥ 18 years	All
Pain	McGill pain questionnaire	≥ 12 years	All
	Wong-Baker Faces Pain rating scale	≥ 2 years	All
Fatigue	Checklist Individual Strength (CIS)	≥ 18 years	All
	Pediatric Quality of Life Inventory (PedsQL) (Multidimensional Fatigue Scale; Child Self-report and/or parent proxy)	2–17 years	All
Activities and participation	ACTIVLIM	≥ 6 years	All
	Impact on Participation and Autonomy (IPA)	≥ 18 years	All
	Egen Klassifikation version 2 (EK2)	≥ 12 years	All
	Borg Rating Scale of Perceived Exertion; prior to and after 6MWT	≥ 5 years	All
Imaging			
Muscles	Muscle ultrasound	All	All
	Full body muscle magnetic resonance imaging (MRI)	≥ 10 years	1, 3
Spine	X-ray of total spine (anteroposterior, lateral) and of lumbar spine (flexion and extension))	≥ 2 years	1, 3
Bone density	Dual-energy X-ray absorptiometry (DEXA-)scan	≥ 2 years	1, 3

Table 1 Examinations performed in LAST STRONG study (Continued)

Outcome domain	Examination	Age	Visit
Cardiopulmonary assessment			
Heart	Electrocardiogram	≥ 2 years	1, 3
	Conventional echocardiography	≥ 2 years	1, 3
Lungs	Spirometry (forced expiratory volume in the first second (FEV1), forced vital capacity (FVC), vital capacity (VC), peak cough flow (PCF))	≥ 5 years	All
	Maximum Expiratory Pressure (MEP), Maximum Inspiratory Pressure (MIP) and Sniff Nasal Inspiratory Pressure (SNIP)	≥ 5 years	All
	Ultrasound of the diaphragm	All	All
Accelerometry			
GENEActiv	Accelerometry for eight consecutive days	≥ 2 years	All

2. Hammersmith Infant Neurological Examinations (HINE) (age < 2 years) [45].
 - a. HINE is designed to be a simple and scorable method for evaluating infants from 2 months to 2 years of age. It includes three sections that assess different aspects of neurologic function, including neurological examination, developmental milestones and behavioral assessment.
3. Motor Function Measure – 20/32 (MFM-20/32) (age ≥ 2 years) [46, 47].
 - a. Motor function in patients with neuromuscular diseases can be measured with the MFM. The MFM is a scale which consists of 20 or 32 items in three dimensions: D1: standing position and transfers, D2: axial and proximal motor function, D3: distal motor function. MFM-32 is used in adults and in children of 7 years and older. Children with the age of 2 to 7 years will undergo MFM-20.
4. Hammersmith Functional Motor Scale (HFMS) (age ≥ 2 years; non-ambulant participants only) [48, 49].
 - a. The HFMS was originally developed to assess the physical abilities of children with non-ambulant spinal muscular atrophy. It consists of 20 items that were considered as important to measure the physical functioning of those patients.
5. Pediatric Balance Scale (PBS) (pediatric patients aged 2–15 years; ambulant participants only) [50].
 - a. The PBS is a modified version of the Berg Balance Scale that is used to assess functional balance skills in school-aged children with mild to moderate motor impairments.
6. Mini Balance Evaluation Systems Test (miniBEST) (age ≥ 16 years; ambulant participants only) [51].
 - a. The miniBEST evaluates balance control by scoring of exercises that belong to one of the following categories: anticipatory postural

changes, reactive postural control, sensory orientation and walking.

7. Graded and timed function tests (age ≥ 2 years: 30 s sit to stand, climb 4 stairs, rise from the floor, timed up and go (TUG); age ≥ 5 years: 6-Minute Walk test, 10-Meter Walk test; ambulant participants only) [52–54].
8. Functional Ambulation Classification (FAC) (age ≥ 5 years) [55].
 - a. The FAC assesses functional ambulation in patients.
9. Brooke and Vignos scale (age ≥ 2 years) [56–58].
 - a. The Brooke and Vignos scales provide ordinal-level data to assess the upper and lower extremity functions, respectively.

Neurological examination and functional measurements will be performed at all four visits.

Questionnaires

Each visit, patients and/or their parent(s) will be asked to complete age-adapted questionnaires on quality of life, pain, fatigue, and activities and participation. These questionnaires include:

1. Pediatric Quality of Life Inventory (PedsQL) (Generic Core Scale, Neuromuscular Module, Multidimensional Fatigue Scale; Child Self-report and/or parent proxy) (pediatric patients 2–17 years) [59–61].
 - a. The PedsQL Generic Core Scale consists of 23 questions in four domains: Physical, Emotional, Social, and School Functioning. It has been translated and subsequently validated into many languages, including Dutch.
 - b. The PedsQL Neuromuscular Module consists of 25 questions in three domains: Neuromuscular disease, Communication and Family resources
 - c. The PedsQL Multidimensional Fatigue Scale assesses subjective fatigue in three domains,

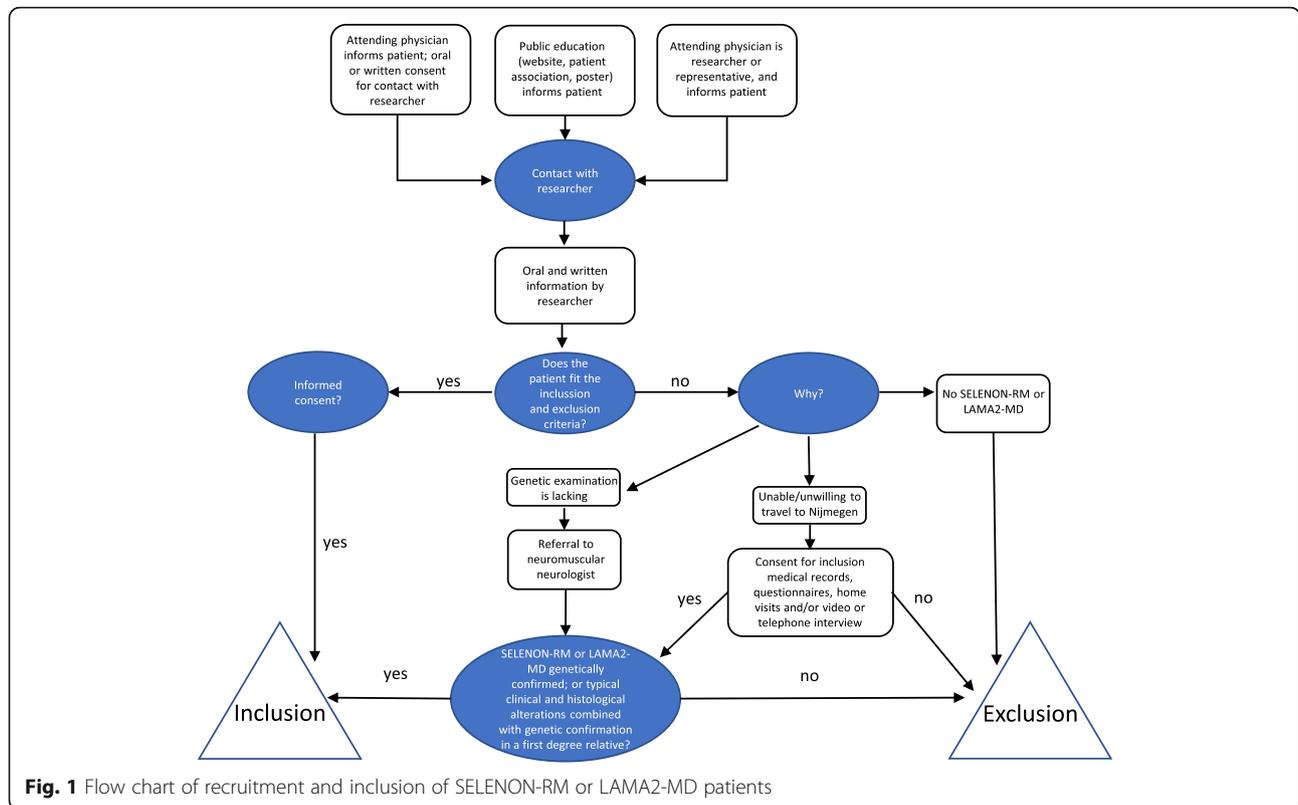


Fig. 1 Flow chart of recruitment and inclusion of SELENON-RM or LAMA2-MD patients

- namely General Fatigue Scale, Sleep/Rest Fatigue Scale, and Cognitive Fatigue Scale.
- 2. Research and Development-36 (RAND36) (age ≥ 18 years) [62].
 - a. Measure for quality of life with 36 items.
- 3. Individualized Neuromuscular Quality of Life (INQoL) (age ≥ 18 years) [63].
 - a. The INQoL is a validated muscle disease specific measure of quality of life, which can be used for individuals or large samples.
- 4. McGill pain questionnaire (age ≥ 12 years) [64].
 - a. Questionnaire in which the location, level and characteristics of pain are assessed.
- 5. Wong-Baker Faces Pain rating scale (age ≥ 2 years) [65].
 - a. The Wong-Baker Faces Pain Scale was originally created for children to help them communicate about their pain.
- 6. Checklist Individual Strength (CIS) (age ≥ 18 years) [66, 67].
 - a. The CIS is a questionnaire rating four subscales: subjective tiredness, concentration, motivation and physical activity. It consists of 20 items on a seven-point scale.
- 7. ACTIVLIM (age ≥ 6 years) [68].
 - a. Questionnaire to assess the ability to perform 22 activities of daily life on a three-point scale from impossible to easy.

- 8. Impact on Participation and Autonomy (IPA) (age ≥ 18 years) [69].
 - a. Questionnaire about participation and autonomy in daily life.
- 9. Egen Klassifikation version 2 (EK2) (age ≥ 12 years and sufficient understanding of the English language) [70].
 - a. The EK2 is a questionnaire that was designed to measure functional ability of activities in daily living in non-ambulant Duchenne muscular dystrophy patients. This questionnaire is available in English. Therefore, only patients who have a sufficient understanding of the English language will be asked to complete this questionnaire.
- 10. Borg Rating Scale of Perceived Exertion (Borg RPE scale) (age ≥ 5 years) [71].
 - a. The Borg RPE scale is used to assess physical activity intensity level. In the LAST STRONG study participants are asked to assess their psychical sensations prior to and after the 6MWT.

Imaging

At all four visits, muscle thickness and muscle echogenicity (quantitative grayscale analysis and Heckmatt ultrasound score) of a subset of bilateral muscles (See Table 2) will be assessed by muscle ultrasound using an

Table 2 Muscle ultrasound in a large subset of bilateral skeletal muscles

Muscle	Point of measurement
M. Temporalis	Parallel to the oculus (above os zygomaticum)
M. Sternocleidomastoid	1/2 line lobulus auricularae to clavícula
M. Biceps brachii	2/3 line acromion – elbow fossa
M. Flexor carpi radialis	1/3 line elbow fossa – caput radii
M. Erector spinae thoracalis	At the level of seventh thoracic vertebrae
M. Erector spinae lumbalis	At the level of third lumbar vertebrae
M. Rectus abdominis	2 cm above umbilicus
M. Biceps femoris	1/2 line gluteal sulcus – popliteal fossa
M. Rectus femoris	1/2 line spina iliaca – upper edge of patella
M. Vastus lateralis	2/3 lateral line spina iliaca – upper edge of patella
M. Gastrocnemius - caput mediale	1/3 line popliteal fossa – medial malleolus
M. Soleus	The place where gastrocnemius disappears and the fibula appears
M. Tibialis anterior	1/3 line lower edge patella – lateral malleolus

Esaote MyLabTwice ultrasound scanner (Esaote SpA, Genoa, Italy) with an 3–13 MHz broadband linear transducer and a 53-mm footprint, adhering to a strictly defined and fixed measurement protocol [72–78]. To measure muscle echogenicity, the mean grayscale level within a manually selected region of interest (ROI) in the ultrasound image is calculated using an in-house developed software package in MATLAB (version 2013b, Mathworks, Natick, MA, USA). The muscle echogenicity and thickness are standardized by calculating their z-score: [(measured grayscale level – predicted grayscale level) / standard deviation grayscale]. The predicted grayscale level is calculated using a reference equation including age, length, sex and weight [78, 79]. In addition, all ultrasound images will be visually evaluated using the semi-quantitative Heckmatt grading scale [80]. At visit 1 (t = 0) and visit 3 (t = 12 months), muscle features are additionally qualitatively and (semi-)quantitatively described through full body muscle MRI (1.5 Tesla, Siemens, Erlangen, Germany) in accordance with our locally developed scanning protocol including Dixon vibe and Short-TI Inversion Recovery (STIR) images [81–84]. In accordance with previous studies on quantitative assessment of muscle MRI, the water and fat image of the Dixon sequence will be used to create a fat fraction map using MATLAB according to the following equation: Fat/(Fat + Water) [84]. The fat fraction map will be used to draw ROI (region of interest) per muscle using ImageJ software (ImageJ 1.47v, National Institutes of Health, USA). ROI will be drawn at predefined localization using the localizer sequences. All drawn ROI

will be checked by a second clinician. Muscle cross-sectional area and fat fractions will be calculated per ROI. The qualitative and semi-quantitative assessment will be performed by two independent assessors. It will include assessment of a predefined subset of bilateral muscles (See Table 3), including relative reduction of muscle volume (0: no reduction, 1: mild, reduction of < 30%, 2: moderate, reduction of 30–60%, 3: severe, reduction of > 60%, 4: non-identifiable muscle), fatty

Table 3 Qualitative and semi-quantitative assessment of a large subset of bilateral skeletal muscles through MRI

Lower extremity and pelvic girdle	Upper extremity, shoulder girdle and trunk
m. Extensor digitorum longus	m. Sternocleidomastoideus
m. Flexor digitorum longus	Neck flexor muscles
m. Gastrocnemius medialis	Neck extensor muscles
m. Gastrocnemius lateralis	m. Levator scapulae
m. Soleus	m. Longus colli
m. Tibialis posterior	m. Latissimus dorsi
m. Tibialis anterior	m. Trapezius
m. Vastus intermedius	m. Deltoideus
m. Vastus medialis	m. Rotatorcuff muscles (Subscapularis)
m. Vastus lateralis	m. Pectoralis major
m. Rectus femoris	m. Pectoralis minor
m. Biceps femoris - short head	m. Serratus anterior
m. Biceps femoris - long head	Anterior arm compartment (m. biceps brachii)
m. Semitendinosus	Posterior arm compartment
m. Semimembranosus	Anterior forearm compartment (m. Flexor carpi radialis)
m. Adductor longus	Posterior forearm compartment
m. Adductor brevis	m. Intercostales
m. Adductor magnus	m. Erector thoracalis spinae
m. Gracilis	m. Erector lumbalis spinae
m. Sartorius	m. Quadratus lumborum
m. Tensor fascia latae	Abdominal belt muscles (m. Rectus abdominis)
m. Quadratus femoris	
m. Gluteus maximus	
m. Gluteus medius	
m. Gluteus minimus	
m. Psoas	
m. Iliacus	
m. Piriformis	
m. Obturator internus	
m. Obturator externus	
m. Perineal muscles	
m. Pectineus	
	Head
	m. Temporalis
	m. Masseter
	m. pterygoideus medialis
	m. pterygoideus lateralis
	Tongue

infiltration (Modified Mercuri scale [85–87]), inflammation (absent/present, plus pattern: patchy, diffuse, peripheral/perifascial, central, other), and fibrosis (absent/present, plus pattern: fascial, intramuscular or other). A MRI will only be performed in patients of 10 years or older who are able to lie still for 45 min without respiratory equipment.

At visit 1 ($t = 0$) and visit 3 ($t = 12$ months), an X-ray will be performed to assess deformities of the spine, including Cobb's angle (spinal curvature), pelvic obliquity, coronal and sagittal balance and lumbar flexion and extension [88, 89]. In order to assess bone density, a DEXA-scan of the right femoral neck and lumbar spine, including a vertebral fracture assessment, will be performed at visit 1 ($t = 0$) and visit 3 ($t = 12$ months) by using the Hologic Discovery A Horizon DXA System (S/N 303053 M).

Cardiac assessment

In order to describe the prevalence and progression of cardiac comorbidities and the need for routine cardiac assessment, patients (age ≥ 2 years) will undergo electrocardiogram and conventional transthoracic echocardiography (TTE) with speckle tracking and Tissue Doppler Imaging (TDI) at visit 1 ($t = 0$) and visit 3 ($t = 12$ months). All investigations will be performed by EACVI TTE certified sonographers using commercially available ultrasound systems (Affiniti70 General, Philips Healthcare, Best, the Netherlands for adult participants; or Vivid E9 or Vivid E95, GE Healthcare Ultrasound, Horten, Norway for pediatric participants). Offline analysis will be performed using dedicated software (AGFA Enterprise Imaging Cardiology version 8.1.2, AGFA Healthcare, Mortsel, Belgium). Global Longitudinal strain (GLS) will be measured using speckle tracking echocardiography on a three beats acquisition with a frame rate > 60 frames/sec. All measurements will be done according to the EACVI recommendations for cardiac chamber quantification [90].

Pulmonary function

At all visits, patients (age ≥ 5 years) will undergo spirometry (forced expiratory volume in the first second (FEV1), forced vital capacity (FVC), vital capacity (VC), peak cough flow (PCF)) in upright and supine position (SpiroUSB, Vyaire Medical connected to PC Spirometry software, Spida CareFusion 2.3.0.10 for Windows 7). Additionally, patients will undergo Maximum Expiratory Pressure (MEP), Maximum Inspiratory Pressure (MIP) and Sniff Nasal Inspiratory Pressure (SNIP) assessment in upright position (Micro RPM, Micro Medical, CareFusion, United Kingdom) [91]. Diaphragm ultrasound (MyLabTwice, Esaote SpA, Genoa, Italy) will be performed to assess diaphragm thickness (end expiratory

and maximum inspiratory thickness) and thickening, and diaphragm echogenicity [92–94].

Accelerometry

After all visits, patients (age ≥ 2 years) are asked to wear an accelerometer (GENEActiv Original, Activinsights Ltd) for eight consecutive days. In the same time period, patients or their parents are asked to fill in a diary with their major activities, including sleeping hours, physical exercise, work and school. The GENEActiv is a tri-axial, wrist-worn accelerometer that will be set to measure at 87,5 Hz sampling [95–97]. The raw data will be converted into 1-s epochs by using the GENEActiv Software (v.3.3, 2019). We will use the gravity subtracted sum of vector magnitudes (SVMgs) as the activity measure. The SVMgs will be measured using the following equation: $SVMgs = \sqrt{(x^2 + y^2 + z^2)} - 1$ g. All data will be subsequently analyzed using MATLAB R2018a Update 4 (9.4.0.902940) for windows. A large subset of parameters will be addressed, including total activity (counts/day), the percentage of sedentary, light, moderate or vigorous activity, and total activity during sleep.

Statistical methods

Due to the explorative character of our study, we will use descriptive statistics (mean, median, SD, 95%-CI) in order to summarize our data. Further, Spearman's correlation analysis and non-parametrical testing will be used to test the correlation between key parameters (i.e. age, ancillary investigations). Further, parameters will be corrected for genetic differences and partial or complete laminin subunit $\alpha 2$ deficiency. If reference values are available from literature, we will check for overlapping confidence intervals. In order to assess disease progression between subsequent measuring moments, we will perform the Wilcoxon signed-rank test (nonparametric continuous paired data) and the McNemar's test (categorical paired data). Multiple linear regressions will be used to explore the relationship between potential disease modifying variables and disease severity. Linear mixed models will be applied for analysis of differences in disease progression. Further, we will correct for multiple testing. We regard $p < 0,05$ as statistically significant. The Statistical Package for the Social Sciences (SPSS version 25, IBM, Armonk, New York) will be used to conduct all statistical analyses.

Data collection

All data-management and data-monitoring will be performed within the Castor software (Version 2021) through direct entry or indirect entry via our electronic patient system (Epic, version May 2020).

Selection of outcome measures for future clinical trials

The results of our natural history study on both SELENON-RM and LAMA2-MD will feed an international key-opinion-leader workshop in which the in- and exclusion criteria, follow-up frequency and outcome measures for the international natural history studies will be discussed. When the results of preclinical and clinical work on promising new therapies continue to be encouraging, a clinical trial will be initiated.

Time plan

Patients will be included between August 2020 and August 2021 and will be followed for 1,5 year. The last visit of the last included patient is expected to take place in the beginning of 2023.

Discussion

Here we present the design of the LAST STRONG study, an extensive natural history study in an unselected cohort of Dutch-speaking SELENON-RM and LAMA2-MD patients (both children and adults) that includes a broad plethora of clinical and functional outcome measures. This enables us to assess the clinical spectrum of patients diagnosed with SELENON-RM or LAMA2-MD. Hereby, we aim to fulfill our primary objectives namely, 1. to assess 1.5-year natural history in patients with SELENON-RM or LAMA2-MD; and 2. to select relevant and sensitive clinical and functional outcome measures to reach trial readiness. In addition, our insights will be vital for reaching our secondary objectives, including making a prevalence estimation, establishing a well-characterized baseline cohort and providing adequate symptomatic management leading to improved clinical care. We propose elaborate qualitative and quantitative measurements adapted to the age and functional abilities of the participant. The structured approach enables us to give an explorative, full-spectrum clinical description of patients diagnosed with SELENON-RM or LAMA2-MD. Further, there is a well-organized health system with full access to (pediatric) neurology and rehabilitation for every patient. Our center also has a longstanding history of patient recruitment for scientific research. Altogether, this is expected to reduce selection bias. Further, as a tertiary referral center for neuromuscular diseases, we are widely experienced with the clinical care and the conduction of studies in patients with rare neuromuscular diseases. Finally, all participants can reach our study center within a three-hour travel by car, which enables us to execute a population based, nationwide study. A consequence of our natural history study might be that the most severely affected patients may be less likely to participate in our study. We expect to overcome this problem to some extent by requesting all available medical records, sending out questionnaires

and performing a medical history by video call/telephone or home visits for all patients that are not able or do not wish to visit our center, provided that written informed consent is given. Due to nationwide travel restrictions and local hospital measures related to the COVID-19 pandemic, Belgian patients are limited in their participation in our study.

The rarity of SELENON-RM and LAMA2-MD will result in a relatively small sample size of Dutch-speaking patients, which limits our possibilities to verify correlations or differences in subgroups. Consequently, we aim to contribute to international collaborations in order to enlarge the availability of natural history data.

The importance to reach trial readiness is warranted by the recent development of promising new treatment strategies for SELENON-RM and LAMA2-MD. Outcome measures need to be reliable and specified per age and disease severity for an adequate measurement of disease progression. If these outcome measures are not reliable, possible positive effects of future treatments in clinical trials will remain to be unrevealed. Reliable outcome measures are thus required in order to start clinical trials.

Further, deep phenotyping of our cohort provides us with valuable information on disease characteristics, enabling us to fulfil one of our secondary objectives, i.e. improving clinical care. For example, participants in whom clinically significant (co)morbidities are found during our study, are referred to the appropriate medical specialists. Moreover, we can take these (co)morbidities into account for routine clinical care of patients with SELENON-RM or LAMA2-MD that are not participating in our natural history study.

In our study, we do not take into account the selection of serum or urine biomarkers in order to limit the burden for patients participating in this study. However, in a study by Bharucha-Goebel et al. serum samples of patients with LAMA2-MD were analyzed for biomarkers. Proteins that were found to be altered, primarily consisted of cytokines and proteins involved in development and cell adhesion. Of the proteins decreased, most were cytokines, growth factors, and protease inhibitors [98]. We recommend to take serum or urine biomarkers into account in future studies.

Conclusion

The LAST STRONG study is expected to provide natural history data, which can be used for the selection of relevant clinical and functional outcome measures in order to reach trial readiness in SELENON-RM and LAMA2-MD patients. Further, our study aims to optimize clinical management in patients diagnosed with SELENON-RM or LAMA2-MD.

Abbreviations

Borg RPE: Borg Rating Scale of Perceived Exertion; CHOP INTEND: The Children's Hospital of Philadelphia Infant Test of Neuromuscular Disorders; CIS: Checklist Individual Strength; DEXA-scan: Dual Energy X-ray absorptiometry; EK2: Egen Klassifikation version 2; FAC: Functional Ambulation Classification; FEV1: Forced expiratory volume in the first second; FVC: Forced vital capacity; HFMS: Hammersmith functional motor scale; HHD: Hand-Held dynamometry; HINE: Hammersmith neurological examinations; INQoL: Individualized Neuromuscular Quality of Life; IPA: Impact on Participation and Autonomy; LAMA2-MD: LAMA2-related muscular dystrophy; MDC1A: Merosin-deficient congenital muscular dystrophy type 1A; MEP: Maximum Expiratory Pressure; MFM-20/32: Motor function measure – 20/32; miniBEST: Mini Balance Evaluation Systems Test; MIP: Maximum Inspiratory Pressure; MRC: Medical Research Council; MRI: Magnetic resonance imaging; PBS: Pediatric Balance Scale; PCF: Peak cough flow; PedsQL: Pediatric Quality of Life Inventory; PROM: Passive range of motion; RAND-36: Reasearch and Development-36; ROI: Region of interest; SELENON-RM: SELENON-related myopathy; SNIP: Sniff Nasal Inspiratory Pressure; STIR: Short-TI Inversion Recovery; SVMgs: The gravity subtracted sum of vector magnitudes; TDI: Tissue doppler imaging; TTE: Transthoracic echocardiography; VC: Vital capacity; 6MWT: 6-Minute Walk Test; 10MWT: 10-Meter Walk Test

Acknowledgements

We thank all patients and their relatives for participation in our study. Several authors of this publication are members of the Netherlands Neuromuscular Center (NL-NMD) and the European Reference Network for rare neuromuscular diseases (EURO-NMD).

Authors' contributions

KB, JTG, JD, NA, FEAU, FMAH, RN, WCMT, SCFMB, ATMD, JMTD, MCHJ, EK, ESBK, SK, JAMS, BK, FHJT, HJMS, BGME, CEE and NCV contributed to the study conception and design. Further, KB, JTG, CEE, NCV drafted the manuscript or revised it critically. All authors have read and approved the final manuscript and agreed to be accountable for their contributions.

Funding

The study is financially supported by a competitively awarded, peer-reviewed grant from Stichting Spieren voor Spieren, Stichting Stofwisselkracht and Stichting Voor Sara, The Netherlands. The scientific advisory boards of the aforementioned foundations took into account the scientific quality of the study protocol, impact of the study, quality of the applicants and requested support. The funders have no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. The content is solely the responsibility of the authors and does not necessarily represent the official views of the Stichting Spieren voor Spieren, Stichting Stofwisselkracht and Stichting Voor Sara.

Availability of data and materials

The datasets generated during the current study will be available in the Donders Repository (<https://data.donders.ru.nl>).

Declarations

Ethics approval and consent to participate

This study is performed in accordance with the Declaration of Helsinki and has been approved by the medical ethical reviewing committee of Region Arnhem-Nijmegen (NL-number NL64269.091.17, dossier number 2017–3911; date of approval of last amendment: 8 July 2020). From all patients, or in case of children their parents or legal guardian, written informed consent will be obtained for participating in our study.

Consent for publication

Not applicable.

Competing interests

Professor Jan A.M. Smeitink is Chief Executive Officer of Khondrion. All other authors declare that they have no competing interests.

Author details

¹Department of Neurology, Donders Institute for Brain, Cognition and Behaviour, Radboud university medical center, Nijmegen, The Netherlands. ²Department of Pediatric Neurology, Donders Institute for Brain, Cognition and Behaviour, Amalia Children's Hospital, Radboud university medical center, Nijmegen, The Netherlands. ³Department of Rehabilitation, Donders Institute for Brain, Cognition and Behaviour, Radboud university medical center, Nijmegen, The Netherlands. ⁴Department of Pediatric cardiology, Amalia Children's Hospital, Radboud university medical center, Nijmegen, The Netherlands. ⁵Department of Cardiology, Radboud university medical center, Nijmegen, The Netherlands. ⁶Department of Radiology, Radboud university medical center, Nijmegen, The Netherlands. ⁷Department of Pediatrics, Amalia Children's Hospital, Radboud university medical center, Nijmegen, The Netherlands. ⁸Department of Internal Medicine, Radboud university medical center, Nijmegen, The Netherlands. ⁹Department of Human Genetics, Radboud university medical center, Nijmegen, The Netherlands. ¹⁰Department of Clinical Genetics, Leiden University Medical Center, Leiden, The Netherlands. ¹¹Khondrion BV, Nijmegen, The Netherlands. ¹²Department of Pathology, Radboud university medical center, Nijmegen, The Netherlands. ¹³Department of Toxicogenomics, Maastricht University, Maastricht, The Netherlands. ¹⁴School for Mental Health and Neurosciences (MHeNS), Maastricht University, Maastricht, the Netherlands. ¹⁵School for Developmental Biology and Oncology (GROW), Maastricht University, Maastricht, The Netherlands.

Received: 23 June 2021 Accepted: 27 July 2021

Published online: 12 August 2021

References

- Witting N, Werlauff U, Duno M, Vissing J. Phenotypes, genotypes, and prevalence of congenital myopathies older than 5 years in Denmark. *Neurol Genet.* 2017;3(2):e140. <https://doi.org/10.1212/NXG.0000000000000140>.
- Villar-Quiles RN, von der Hagen M, Métaiz C, Gonzalez V, Donkervoort S, Bertini E, et al. The clinical, histologic, and genotypic spectrum of. *Neurology.* 2020;95(11):e1512–e27. <https://doi.org/10.1212/WNL.00000000000010327>.
- Nguyen Q, Lim KRQ, Yokota T. Current understanding and treatment of cardiac and skeletal muscle pathology in laminin- α 2 chain-deficient congenital muscular dystrophy. *Appl Clin Genet.* 2019;12:113–30. <https://doi.org/10.2147/TACG.S187481>.
- Geranmayeh F, Clement E, Feng LH, Sewry C, Pagan J, Mein R, et al. Genotype-phenotype correlation in a large population of muscular dystrophy patients with LAMA2 mutations. *Neuromuscul Disord.* 2010;20(4):241–50. <https://doi.org/10.1016/j.nmd.2010.02.001>.
- Sarkozy A, Foley AR, Zambon AA, Bönnemann CG, Muntoni F. LAMA2-related dystrophies: clinical phenotypes, disease biomarkers, and clinical trial readiness. *Front Mol Neurosci.* 2020;13:123. <https://doi.org/10.3389/fnmol.2020.00123>.
- Moghadaszadeh B, Petit N, Jaillard C, Brockington M, Quijano Roy S, Merlini L, et al. Mutations in SEPN1 cause congenital muscular dystrophy with spinal rigidity and restrictive respiratory syndrome. *Nat Genet.* 2001;29(1):17–8. <https://doi.org/10.1038/ng713>.
- Ferreiro A, Quijano-Roy S, Pichereau C, Moghadaszadeh B, Goemans N, Bönnemann C, et al. Mutations of the selenoprotein N gene, which is implicated in rigid spine muscular dystrophy, cause the classical phenotype of multiminicore disease: reassessing the nosology of early-onset myopathies. *Am J Hum Genet.* 2002;71(4):739–49. <https://doi.org/10.1086/342719>.
- Clarke NF, Kidson W, Quijano-Roy S, Estournet B, Ferreiro A, Guicheney P, et al. SEPN1: associated with congenital fiber-type disproportion and insulin resistance. *Ann Neurol.* 2006;59(3):546–52. <https://doi.org/10.1002/ana.20761>.
- Ferreiro A, Ceuterick-de Groote C, Marks JJ, Goemans N, Schreiber G, Hanefeld F, et al. Desmin-related myopathy with Mallory body-like inclusions is caused by mutations of the selenoprotein N gene. *Ann Neurol.* 2004;55(5):676–86. <https://doi.org/10.1002/ana.20077>.
- Wang CH, Bönnemann CG, Rutkowski A, Sejersen T, Bellini J, Battista V, et al. Consensus statement on standard of care for congenital muscular dystrophies. *J Child Neurol.* 2010;25(12):1559–81. <https://doi.org/10.1177/0883073810381924>.

11. Smeets HJM, Verbrugge B, Springuel P, Voermans NC, group MAW. International Workshop Report Congenital muscular dystrophy 1A: the road to therapy. *Neuromuscul Disord*. 2021.
12. Janssen MCH, Koene S, de Laat P, Hemelaar P, Pickkers P, Spaans E, et al. The KHENERGY study: safety and efficacy of KH176 in mitochondrial m. 3243A>G Spectrum disorders. *Clin Pharmacol Ther*. 2019;105(1):101–11. <https://doi.org/10.1002/cpt.1197>.
13. Moulin M, Ferreiro A. Muscle redox disturbances and oxidative stress as pathomechanisms and therapeutic targets in early-onset myopathies. *Semin Cell Dev Biol*. 2017;64:213–23. <https://doi.org/10.1016/j.semcdb.2016.08.003>.
14. Arbogast S, Beuvin M, Fraysse B, Zhou H, Muntoni F, Ferreiro A. Oxidative stress in SEPN1-related myopathy: from pathophysiology to treatment. *Ann Neurol*. 2009;65(6):677–86. <https://doi.org/10.1002/ana.21644>.
15. Arbogast S, Ferreiro A. Selenoproteins and protection against oxidative stress: selenoprotein N as a novel player at the crossroads of redox signaling and calcium homeostasis. *Antioxid Redox Signal*. 2010;12(7):893–904. <https://doi.org/10.1089/ars.2009.2890>.
16. Filipe A, Chernorudskiy A, Arbogast S, Varone E, Villar-Quiles RN, Pozzer D, et al. Defective endoplasmic reticulum-mitochondria contacts and bioenergetics in SEPN1-related myopathy. *Cell Death Differ*. 2021;28(1):123–38. <https://doi.org/10.1038/s41418-020-0587-z>.
17. Pozzer D, Varone E, Chernorudskiy A, Schiarea S, Missiroli S, Giorgi C, et al. A maladaptive ER stress response triggers dysfunction in highly active muscles of mice with SELENON loss. *Redox Biol*. 2019;20:354–66. <https://doi.org/10.1016/j.redox.2018.10.017>.
18. Kemaladewi DU, Bassi PS, Erwood S, Al-Basha D, Gawlik KI, Lindsay K, et al. A mutation-independent approach for muscular dystrophy via upregulation of a modifier gene. *Nature*. 2019;572(7767):125–30. <https://doi.org/10.1038/s41586-019-1430-x>.
19. Reinhard JR, Lin S, McKee KK, Meinen S, Crosson SC, Sury M, et al. Linker proteins restore basement membrane and correct LAMA2-related muscular dystrophy in mice. *Sci Transl Med*. 2017;9(396).
20. Rooney JE, Knapp JR, Hodges BL, Wuebbles RD, Burkin DJ. Laminin-111 protein therapy reduces muscle pathology and improves viability of a mouse model of merosin-deficient congenital muscular dystrophy. *Am J Pathol*. 2012;180(4):1593–602. <https://doi.org/10.1016/j.ajpath.2011.12.019>.
21. Barraza-Flores P, Bates CR, Oliveira-Santos A, Burkin DJ. Laminin and integrin in LAMA2-related congenital muscular dystrophy: from disease to therapeutics. *Front Mol Neurosci*. 2020;13:1. <https://doi.org/10.3389/fnmol.2020.00001>.
22. Chernorudskiy A, Varone E, Colombo SF, Fumagalli S, Cagnotto A, Cattaneo A, et al. Selenoprotein N is an endoplasmic reticulum calcium sensor that links luminal calcium levels to a redox activity. *Proc Natl Acad Sci U S A*. 2020;117(35):21288–98. <https://doi.org/10.1073/pnas.2003847117>.
23. Patton BL, Miner JH, Chiu AY, Sanes JR. Distribution and function of laminins in the neuromuscular system of developing, adult, and mutant mice. *J Cell Biol*. 1997;139(6):1507–21. <https://doi.org/10.1083/jcb.139.6.1507>.
24. Petrof BJ, Shrager JB, Stedman HH, Kelly AM, Sweeney HL. Dystrophin protects the sarcolemma from stresses developed during muscle contraction. *Proc Natl Acad Sci U S A*. 1993;90(8):3710–4. <https://doi.org/10.1073/pnas.90.8.3710>.
25. Fontes-Oliveira CC, Steinz M, Schneiderat P, Mulder H, Durbeef M. Bioenergetic impairment in congenital muscular dystrophy type 1A and Leigh syndrome muscle cells. *Sci Rep*. 2017;7(1):45272. <https://doi.org/10.1038/srep45272>.
26. Harandi VM, Oliveira BMS, Allamand V, Friberg A, Fontes-Oliveira CC, Durbeef M. Antioxidants Reduce Muscular Dystrophy. *Antioxidants*. 2020;9(3).
27. Kemaladewi D, Hyatt E, Ivakine Z, Cohn R. CRISPR/Cas9-mediated exon inclusion in Lama2 gene alleviates dystrophic pathology in MDC1A mouse model in title abstract keyword. *Neuromuscul Disord*. 2016;26:S190. <https://doi.org/10.1016/j.nmd.2016.06.376>.
28. Wright M. Calcium handling in a zebrafish model of SELENON congenital muscular dystrophy. *World Muscle Society: Neuromuscular Disorders*; 2020. p. S149–S50.
29. Deniziak M, Thisse C, Rederstorff M, Hindelang C, Thisse B, Lescure A. Loss of selenoprotein N function causes disruption of muscle architecture in the zebrafish embryo. *Exp Cell Res*. 2007;313(1):156–67. <https://doi.org/10.1016/j.yexcr.2006.10.005>.
30. Bachmann C, Noreen F, Voermans NC, Schär PL, Vissing J, Fock JM, et al. Aberrant regulation of epigenetic modifiers contributes to the pathogenesis in patients with selenoprotein N-related myopathies. *Hum Mutat*. 2019;40(7):962–74. <https://doi.org/10.1002/humu.23745>.
31. Marino M, Stoilova T, Giorgi C, Bachi A, Cattaneo A, Auricchio A, et al. SEPN1, an endoplasmic reticulum-localized selenoprotein linked to skeletal muscle pathology, counteracts hyperoxidation by means of redox-regulating SERCA2 pump activity. *Hum Mol Genet*. 2015;24(7):1843–55. <https://doi.org/10.1093/hmg/ddu602>.
32. Yurchenco PD, McKee KK, Reinhard JR, Rüegg MA. Laminin-deficient muscular dystrophy: Molecular pathogenesis and structural repair strategies. *Matrix Biol*. 2018;71–72:174–87.
33. Hara Y, Mizobe Y, Miyatake S, Takizawa H, Nagata T, Yokota T, et al. Exon skipping using antisense oligonucleotides for laminin-Alpha2-deficient muscular dystrophy. *Methods Mol Biol*. 2018;2018:553–64.
34. Hall TE, Wood AJ, Ehrlich O, Li M, Sonntag CS, Cole NJ, et al. Cellular rescue in a zebrafish model of congenital muscular dystrophy type 1A. *NPJ Regen Med*. 2019;4(1):21. <https://doi.org/10.1038/s41536-019-0084-5>.
35. Meilleur KG, Jain MS, Hynan LS, Shieh CY, Kim E, Waite M, et al. Results of a two-year pilot study of clinical outcome measures in collagen VI- and laminin alpha2-related congenital muscular dystrophies. *Neuromuscul Disord*. 2015;25(1):43–54. <https://doi.org/10.1016/j.nmd.2014.09.010>.
36. Jain MS, Meilleur K, Kim E, Norato G, Waite M, Nelson L, et al. Longitudinal changes in clinical outcome measures in COL6-related dystrophies and LAMA2-related dystrophies. *Neurology*. 2019;93(21):e1932–e43. <https://doi.org/10.1212/WNL.0000000000008517>.
37. Zambon AA, Ridout D, Main M, Mein R, Phadke R, Muntoni F, et al. LAMA2-related muscular dystrophy: natural history of a large pediatric cohort. *Ann Clin Transl Neurol*. 2020;7(10):1870–82. <https://doi.org/10.1002/acn3.51172>.
38. Beenakker EA, van der Hoeven JH, Fock JM, Maurits NM. Reference values of maximum isometric muscle force obtained in 270 children aged 4–16 years by hand-held dynamometry. *Neuromuscul Disord*. 2001;11(5):441–6. [https://doi.org/10.1016/S0960-8966\(01\)00193-6](https://doi.org/10.1016/S0960-8966(01)00193-6).
39. van den Beld WA, van der Sanden GA, Sengers RC, Verbeek AL, Gabreëls FJ. Validity and reproducibility of hand-held dynamometry in children aged 4–11 years. *J Rehabil Med*. 2006;38(1):57–64. <https://doi.org/10.1080/16501970510044043>.
40. McKay MJ, Baldwin JN, Ferreira P, Simic M, Vanicek N, Burns J, et al. Normative reference values for strength and flexibility of 1,000 children and adults. *Neurology*. 2017;88(1):36–43. <https://doi.org/10.1212/WNL.0000000000003466>.
41. van der Ploeg RJ, Fidler V, Oosterhuis HJ. Hand-held myometry: reference values. *J Neurol Neurosurg Psychiatry*. 1991;54(3):244–7. <https://doi.org/10.1136/jnnp.54.3.244>.
42. Soucie JM, Wang C, Forsyth A, Funk S, Denny M, Roach KE, et al. Range of motion measurements: reference values and a database for comparison studies. *Haemophilia*. 2011;17(3):500–7. <https://doi.org/10.1111/j.1365-2516.2010.02399.x>.
43. Glanzman AM, Mazzone E, Main M, Pelliccioni M, Wood J, Swoboda KJ, et al. The Children's Hospital of Philadelphia infant test of neuromuscular disorders (CHOP INTEND): test development and reliability. *Neuromuscul Disord*. 2010;20(3):155–61. <https://doi.org/10.1016/j.nmd.2009.11.014>.
44. Glanzman AM, McDermott MP, Montes J, Martens WB, Flickinger J, Riley S, et al. Validation of the Children's Hospital of Philadelphia infant test of neuromuscular disorders (CHOP INTEND). *Pediatr Phys Ther*. 2011;23(4):322–6. <https://doi.org/10.1097/PEP.0b013e3182351f04>.
45. Dubowitz L, Riccio D, Mercuri E. The Dubowitz neurological examination of the full-term newborn. *Ment Retard Dev Disabil Res Rev*. 2005;11(1):52–60. <https://doi.org/10.1002/mrdd.20048>.
46. de Lattre C, Payan C, Vuillerot C, Rippert P, de Castro D, Bérard C, et al. Motor function measure: validation of a short form for young children with neuromuscular diseases. *Arch Phys Med Rehabil*. 2013;94(11):2218–26. <https://doi.org/10.1016/j.apmr.2013.04.001>.
47. Bérard C, Payan C, Hodgkinson J, Fermanian J, Group MCS. A motor function measure for neuromuscular diseases. Construction and validation study. *Neuromuscul Disord*. 2005;15(7):463–70. <https://doi.org/10.1016/j.nmd.2005.03.004>.
48. Main M, Kairon H, Mercuri E, Muntoni F. The Hammersmith functional motor scale for children with spinal muscular atrophy: a scale to test ability and monitor progress in children with limited ambulation. *Eur J Paediatr Neurol*. 2003;7(4):155–9. [https://doi.org/10.1016/S1090-3798\(03\)00060-6](https://doi.org/10.1016/S1090-3798(03)00060-6).
49. Ramsey D, Scoto M, Mayhew A, Main M, Mazzone ES, Montes J, et al. Revised Hammersmith scale for spinal muscular atrophy: a SMA specific clinical outcome assessment tool. *PLoS One*. 2017;12(2):e0172346. <https://doi.org/10.1371/journal.pone.0172346>.

50. Franjoine MR, Gunther JS, Taylor MJ. Pediatric balance scale: a modified version of the berg balance scale for the school-age child with mild to moderate motor impairment. *Pediatr Phys Ther.* 2003;15(2):114–28. <https://doi.org/10.1097/01.PEP.0000068117.48023.18>.
51. Franchignoni F, Horak F, Godi M, Nardone A, Giordano A. Using psychometric techniques to improve the balance evaluation systems test: the mini-BESTest. *J Rehabil Med.* 2010;42(4):323–31. <https://doi.org/10.2340/16501977-0537>.
52. Florence JM, Pandya S, King WM, Robison JD, Signore LC, Wentzell M, et al. Clinical trials in Duchenne dystrophy. Standardization and reliability of evaluation procedures. *Phys Ther.* 1984;64(1):41–5. <https://doi.org/10.1093/ptj/64.1.41>.
53. Mayhew JE, Florence JM, Mayhew TP, Henricson EK, Leshner RT, McCarter RJ, et al. Reliable surrogate outcome measures in multicenter clinical trials of Duchenne muscular dystrophy. *Muscle Nerve.* 2007;35(1):36–42. <https://doi.org/10.1002/mus.20654>.
54. Enright PL. The six-minute walk test. *Respir Care.* 2003;48(8):783–5.
55. Holden MK, Gill KM, Magliozzi MR, Nathan J, Piehl-Baker L. Clinical gait assessment in the neurologically impaired. Reliability and meaningfulness. *Phys Ther.* 1984;64(1):35–40. <https://doi.org/10.1093/ptj/64.1.35>.
56. Jung IY, Chae JH, Park SK, Kim JH, Kim JY, Kim SJ, et al. The correlation analysis of functional factors and age with duchenne muscular dystrophy. *Ann Rehabil Med.* 2012;36(1):22–32. <https://doi.org/10.5535/arm.2012.36.1.22>.
57. Brooke MH, Griggs RC, Mendell JR, Fenichel GM, Shumate JB, Pellegrino RJ. Clinical trial in Duchenne dystrophy. I. the design of the protocol. *Muscle Nerve.* 1981;4(3):186–97. <https://doi.org/10.1002/mus.880040304>.
58. Lue YJ, Lin RF, Chen SS, Lu YM. Measurement of the functional status of patients with different types of muscular dystrophy. *Kaohsiung J Med Sci.* 2009;25(6):325–33. [https://doi.org/10.1016/S1607-551X\(09\)70523-6](https://doi.org/10.1016/S1607-551X(09)70523-6).
59. Engelen V, Haentjens MM, Detmar SB, Koopman HM, Grootenhuys MA. Health related quality of life of Dutch children: psychometric properties of the PedsQL in the Netherlands. *BMC Pediatr.* 2009;9(1):68. <https://doi.org/10.1186/1471-2431-9-68>.
60. Iannaccone ST, Hyman LS, Morton A, Buchanan R, Limbers CA, Varni JW, et al. The PedsQL in pediatric patients with spinal muscular atrophy: feasibility, reliability, and validity of the pediatric quality of life inventory generic Core scales and neuromuscular module. *Neuromuscul Disord.* 2009;19(12):805–12. <https://doi.org/10.1016/j.nmd.2009.09.009>.
61. Gordijn M, Suzanne Gordijn M, Cremers EM, Kaspers GJ, Gemke RJ. Fatigue in children: reliability and validity of the Dutch PedsQL™ multidimensional fatigue scale. *Qual Life Res.* 2011;20(7):1103–8. <https://doi.org/10.1007/s11360-010-9836-9>.
62. Aaronson NK, Muller M, Cohen PD, Essink-Bot ML, Fekkes M, Sanderman R, et al. Translation, validation, and norming of the Dutch language version of the SF-36 health survey in community and chronic disease populations. *J Clin Epidemiol.* 1998;51(11):1055–68. [https://doi.org/10.1016/S0895-4356\(98\)00097-3](https://doi.org/10.1016/S0895-4356(98)00097-3).
63. Seesing FM, van Vught LE, Rose MR, Drost G, van Engelen BG, van der Wilt GJ. The individualized neuromuscular quality of life questionnaire: cultural translation and psychometric validation for the Dutch population. *Muscle Nerve.* 2015;51(4):496–500. <https://doi.org/10.1002/mus.24337>.
64. Vanderiet K, Adriaansen H, Carton H, Vertommen H. The McGill pain questionnaire constructed for the Dutch language (MPQ-DV). Preliminary data concerning reliability and validity. *Pain.* 1987;30(3):395–408. [https://doi.org/10.1016/0304-3959\(87\)90027-3](https://doi.org/10.1016/0304-3959(87)90027-3).
65. Wong DL, Baker CM. Pain in children: comparison of assessment scales. *Pediatr Nurs.* 1988;14(1):9–17.
66. Worm-Smeitink M, Gielissen M, Bloot L, van Laarhoven HWM, van Engelen BGM, van Riel P, et al. The assessment of fatigue: psychometric qualities and norms for the checklist individual strength. *J Psychosom Res.* 2017;98:40–6. <https://doi.org/10.1016/j.jpsychores.2017.05.007>.
67. Vercoulen JH, Swanink CM, Fennis JF, Galama JM, van der Meer JW, Blijenberg G. Dimensional assessment of chronic fatigue syndrome. *J Psychosom Res.* 1994;38(5):383–92. [https://doi.org/10.1016/0022-3999\(94\)90099-X](https://doi.org/10.1016/0022-3999(94)90099-X).
68. Vandervelde L, Van den Bergh PY, Goemans N, Thonnard JL. ACTIVLIM: a Rasch-built measure of activity limitations in children and adults with neuromuscular disorders. *Neuromuscul Disord.* 2007;17(6):459–69. <https://doi.org/10.1016/j.nmd.2007.02.013>.
69. Cardol M, de Haan RJ, van den Bos GA, de Jong BA, de Groot IJ. The development of a handicap assessment questionnaire: the impact on participation and autonomy (IPA). *Clin Rehabil.* 1999;13(5):411–9. <https://doi.org/10.1191/02692159968601325>.
70. Steffensen B, Hyde S, Lyager S, Mattsson E. Validity of the EK scale: a functional assessment of non-ambulatory individuals with Duchenne muscular dystrophy or spinal muscular atrophy. *Physiother Res Int.* 2001;6(3):119–34. <https://doi.org/10.1002/pri.221>.
71. Borg GA. Psychophysical bases of perceived exertion. *Med Sci Sports Exerc.* 1982;14(5):377–81.
72. van Alfen N, Mah JK. Neuromuscular ultrasound: a new tool in your toolbox. *Can J Neurol Sci.* 2018;45(5):504–15. <https://doi.org/10.1017/cjn.2018.269>.
73. Mah JK, van Alfen N. Neuromuscular ultrasound: clinical applications and diagnostic values. *Can J Neurol Sci.* 2018;45(6):605–19. <https://doi.org/10.1017/cjn.2018.314>.
74. Pillen S, van Alfen N. Skeletal muscle ultrasound. *Neurol Res.* 2011;33(10):1016–24. <https://doi.org/10.1179/1743132811Y.0000000010>.
75. Pillen S, van Alfen N. Muscle ultrasound from diagnostic tool to outcome measure—quantification is the challenge. *Muscle Nerve.* 2015;52(3):319–20. <https://doi.org/10.1002/mus.24613>.
76. Pillen S, Verrips A, van Alfen N, Arts IM, Sie LT, Zwartz MJ. Quantitative skeletal muscle ultrasound: diagnostic value in childhood neuromuscular disease. *Neuromuscul Disord.* 2007;17(7):509–16. <https://doi.org/10.1016/j.nmd.2007.03.008>.
77. Pillen S, Boon A, Van Alfen N. Muscle ultrasound. *Handb Clin Neurol.* 2016;136:843–53. <https://doi.org/10.1016/B978-0-444-53486-6.00042-9>.
78. Wijntjes J, van Alfen N. Muscle ultrasound: present state and future opportunities. *Muscle Nerve.* 2021;63(4):455–66. <https://doi.org/10.1002/mus.27081>.
79. Scholten RR, Pillen S, Verrips A, Zwartz MJ. Quantitative ultrasonography of skeletal muscles in children: normal values. *Muscle Nerve.* 2003;27(6):693–8. <https://doi.org/10.1002/mus.10384>.
80. Heckmatt JZ, Leeman S, Dubowitz V. Ultrasound imaging in the diagnosis of muscle disease. *J Pediatr.* 1982;101(5):656–60. [https://doi.org/10.1016/S0022-3476\(82\)80286-2](https://doi.org/10.1016/S0022-3476(82)80286-2).
81. Dahlqvist JR, Widholm P, Leinhard OD, Vissing J. MRI in neuromuscular diseases: an emerging diagnostic tool and biomarker for prognosis and efficacy. *Ann Neurol.* 2020;88(4):669–81. <https://doi.org/10.1002/ana.25804>.
82. Morrow JM, Sinclair CD, Fischmann A, Machado PM, Reilly MM, Yousry TA, et al. MRI biomarker assessment of neuromuscular disease progression: a prospective observational cohort study. *Lancet Neurol.* 2016;15(1):65–77. [https://doi.org/10.1016/S1474-4422\(15\)00242-2](https://doi.org/10.1016/S1474-4422(15)00242-2).
83. Mul K, Horlings CGC, Vincenten SCC, Voermans NC, van Engelen BGM, van Alfen N. Quantitative muscle MRI and ultrasound for facioscapulohumeral muscular dystrophy: complementary imaging biomarkers. *J Neurol.* 2018;265(11):2646–55. <https://doi.org/10.1007/s00415-018-9037-y>.
84. Mul K, Vincenten SCC, Voermans NC, Lemmers RJLF, van der Vliet PJ, van der Maarel SM, et al. Adding quantitative muscle MRI to the FSHD clinical trial toolbox. *Neurology.* 2017;89(20):2057–65. <https://doi.org/10.1212/WNL.0000000000004647>.
85. Kinali M, Arechavala-Gomez V, Cirak S, Glover A, Guglieri M, Feng L, et al. Muscle histology vs MRI in Duchenne muscular dystrophy. *Neurology.* 2011;76(4):346–53. <https://doi.org/10.1212/WNL.0b013e318208811f>.
86. Mercuri E, Cini C, Pichiecchio A, Allsop J, Counsell S, Zolkipli Z, et al. Muscle magnetic resonance imaging in patients with congenital muscular dystrophy and Ullrich phenotype. *Neuromuscul Disord.* 2003;13(7–8):554–8. [https://doi.org/10.1016/S0960-8966\(03\)00091-9](https://doi.org/10.1016/S0960-8966(03)00091-9).
87. Mercuri E, Pichiecchio A, Counsell S, Allsop J, Cini C, Jungbluth H, et al. A short protocol for muscle MRI in children with muscular dystrophies. *Eur J Paediatr Neurol.* 2002;6(6):305–7. <https://doi.org/10.1053/ejpn.2002.0617>.
88. Gupta MC, Wijesekera S, Sossan A, Martin L, Vogel LC, Boakes JL, et al. Reliability of radiographic parameters in neuromuscular scoliosis. *Spine (Phila Pa 1976).* 2007;32(6):691–5. <https://doi.org/10.1097/01.brs.0000257524.23074.ed>.
89. Kim H, Kim HS, Moon ES, Yoon CS, Chung TS, Song HT, et al. Scoliosis imaging: what radiologists should know. *Radiographics.* 2010;30(7):1823–42. <https://doi.org/10.1148/rg.307105061>.
90. Lang RM, Badano LP, Mor-Avi V, Afilalo J, Armstrong A, Ernande L, et al. Recommendations for cardiac chamber quantification by echocardiography in adults: an update from the American Society of Echocardiography and the European Association of Cardiovascular Imaging. *Eur Heart J Cardiovasc Imaging.* 2015;16(3):233–70. <https://doi.org/10.1093/ehjci/jev014>.
91. Laveneziana P, Albuquerque A, Aliverti A, Babb T, Barreiro E, Dres M, et al. ERS statement on respiratory muscle testing at rest and during exercise. *Eur Respir J.* 2019;53(6).

92. Fayssoil A, Nguyen LS, Ogna A, Stojkovic T, Meng P, Mompoint D, et al. Diaphragm sniff ultrasound: Normal values, relationship with sniff nasal pressure and accuracy for predicting respiratory involvement in patients with neuromuscular disorders. *PLoS One*. 2019;14(4):e0214288. <https://doi.org/10.1371/journal.pone.0214288>.
93. Caggiano S, Khirani S, Dabaj I, Cavassa E, Amaddeo A, Arroyo JO, et al. Diaphragmatic dysfunction in SEPNI-related myopathy. *Neuromuscul Disord*. 2017;27(8):747–55. <https://doi.org/10.1016/j.nmd.2017.04.010>.
94. van Doorn JLM, Pennati F, Hansen HHG, van Engelen BGM, Aliverti A, Doorduyn J. Respiratory muscle imaging by ultrasound and MRI in neuromuscular disorders. *Eur Respir J*. 2021.
95. Esliger DW, Rowlands AV, Hurst TL, Catt M, Murray P, Eston RG. Validation of the GENEa accelerometer. *Med Sci Sports Exerc*. 2011;43(6):1085–93. <https://doi.org/10.1249/MSS.0b013e31820513be>.
96. Phillips LR, Parfitt G, Rowlands AV. Calibration of the GENEa accelerometer for assessment of physical activity intensity in children. *J Sci Med Sport*. 2013;16(2):124–8. <https://doi.org/10.1016/j.jsams.2012.05.013>.
97. de Vries PR, Janssen M, Spaans E, de Groot I, Janssen A, Smeitink J, et al. Natural variability of daily physical activity measured by accelerometry in children with a mitochondrial disease. *Mitochondrion*. 2019;47:30–7. <https://doi.org/10.1016/j.mito.2019.04.005>.
98. Bharucha-Goebel D, Collins J, Hu Y, Foley AR, Donkervoort S, Leach M, et al. Serum biomarker discovery for congenital muscular dystrophies. *Neuromuscul Disord*. 2016.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

